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(54) Title: CUTINASE VARIANTS

(57) Abstract

Variants of fungal cutinases have improved thermostability. The variants comprise substitution of one or more amino acid residues near the N-terminal in the amino acid sequence or in the three-dimensional structure of the cutinase.

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1

CUTINASE VARIANTS

FIELD OF THE INVENTION

The present invention relates to a cutinase variant, more particularly to a cutinase variant having improved thermostability. The invention also relates to a DNA sequence encoding the variant, a vector comprising the DNA sequence, a transformed host cell harboring the DNA sequence or the vector, to a method of producing the variant, and to use of the variant.

BACKGROUND OF THE INVENTION

Cutinases are lipolytic enzymes capable of hydrolyzing the substrate cutin.

10 Cutinases are known from various fungi (P.E. Kolattukudy in "Lipases", Ed. B. Borgström and H.L. Brockman, Elsevier 1984, 471-504). The amino acid sequence and the crystal structure of a cutinase of *Fusarium solani pisi* have been described (S. Longhi et al., Journal of Molecular Biology, 268 (4), 779-799 (1997)). The amino acid sequence of a cutinase from *Humicola insolens* has also been published (US 5,827,719).

A number of variants of the cutinase of *Fusarium solani pisi* have been published: WO 94/14963; WO 94/14964; Appl. Environm. Microbiol. 64, 2794-2799, 1998; Proteins: Structure, Function and Genetics 26, 442-458, 1996; J. of Computational Chemistry 17, 1783-1803, 1996; Protein Engineering 6, 157-165, 1993; Proteins: Structure, Function, and Genetics 33, 253-264, 1998; J. of Biotechnology 66, 11-26, 1998; Biochemistry 35, 398-410, 1996.

Fungal cutinases may be used in the enzymatic hydrolysis of cyclic oligomers of poly(ethylene terephthalate), e.g. in the finishing of yarn or fabric from poly(ethylene terephthalate) fibers (WO 97/27237). However, it is desirable to improve the thermostability of known fungal cutinases to allow a higher process temperature.

SUMMARY OF THE INVENTION

The inventors have found certain variants of fungal cutinases having improved thermostability.

2

Accordingly, the invention provides a variant of a parent fungal cutinase comprising substitution of one or more amino acid residues which is located:

- a) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
- b) within 20 positions from the N-terminal amino acid.

The invention also provides a DNA sequence encoding the variant, an expression vector comprising the DNA sequence, a transformed host cell harboring the DNA sequence or the expression vector, a method of producing the variant, processes using the variant and a detergent composition comprising the variant.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 gives the coordinates for the 3D structure of the cutinase of *H. insolens*.

Fig. 2 is a computer model showing the three-dimensional structures of the cutinases from *F. solani pisi* (left) and *H. insolens* (right). Different colors have been used to identify the N-terminal amino acid and zones of 12 Å and 17 Å diameter around this.

Figs. 3-6 illustrate the hydrolysis of c3ET. Details are given in the Examples.

DETAILED DESCRIPTION OF THE INVENTION

20 Fungal cutinase

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The parent cutinase is a fungal cutinase, such as a filamentous fungal cutinase, e.g. native to a strain of *Humicola* or *Fusarium*, specifically *H. insolens* or *F. solani pisi*, more specifically *H. insolens* strain DSM 1800.

The amino acid sequence of the cutinase of *H. insolens* strain DSM 1800 and the DNA sequence encoding it are shown as SEQ ID NO: 2 and SEQ ID NO: 1 of US 5,827,719. The numbering system used herein for the *H. insolens* cutinase is based on the mature peptide, as shown in said SEQ ID NO: 2.

The amino acid sequence of the cutinase of *F. solani pisi* is shown as the mature peptide in Fig. 1D of WO 94/14964. The numbering system used herein for

3

the *F. solani pisi* cutinase is that used in WO 94/14964; it includes the prosequence shown in said Fig. 1D; thus, the mature cutinase is at positions 16-214.

The parent cutinase may have an amino acid sequence which is at least 50 % (particularly at least 70 % or at least 80 %) homologous to the cutinase of *H. insolens* strain DSM 1800. The parent cutinase may particularly be one that can be aligned with the cutinase of *H. insolens* strain DSM 1800.

Nomenclature for amino acids and alterations

The specification and claims refer to amino acids by their one-letter codes. A particular amino acid in a sequence is identified by its one-letter code and its position, e.g. Q1 indicates Gln (glutamine at position 1, i.e. at the N-terminal.

The nomenclature used herein for defining substitutions is basically as described in WO 92/05249. Thus, R51P indicates substitution of R (Arg) at position 51 with P (Pro).

Homology and alignment

For purposes of the present invention, the degree of homology may be suitably determined according to the method described in Needleman, S.B. and Wunsch, C.D., (1970), Journal of Molecular Biology, 48, 443-45, with the following settings for polypeptide sequence comparison: GAP creation penalty of 3.0 and GAP extension penalty of 0.1. The determination may be done by means of a computer program known such as GAP provided in the GCG program package (Program Manual for the Wisconsin Package, Version 8, August 1994, Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA 53711).

Two given sequences can be aligned according to the method described in Needleman (*supra*) using the same parameters. This may be done by means of the GAP program (*supra*).

Three-dimensional structure of cutinase

The structure of the cutinase of *H. insolens* was solved in accordance with the principle for X-ray crystallographic methods as given, for example, in X-Ray Structure Determination, Stout, G.K. and Jensen, L.H., John Wiley & Sons, Inc. NY, 30 1989. The structural coordinates for the solved crystal structure at 2.2 Å resolution

4

using the isomorphous replacement method are given in Fig. 1 in standard PDB format (Protein Data Bank, Brookhaven National Laboratory, Brookhaven, CT).

The structure of the cutinase of *F. solani pisi* is described in Martinez et al. (1992) Nature 356, 615-618. The 3D structures of the cutinases of *F. solani pisi* and *H. insolens* are compared as a computer model in Fig. 2.

It should be noted that the overall three-dimensional structures of fungal cutinases are very similar and have been shown by X-ray crystallography to be highly homologous. The similarities between the cutinases from *F. solani pisi* and *H. insolens* are clearly apparent from the computer model in Fig. 2. Therefore, modifications of the type indicated for one fungal cutinase will also be functional for other fungal cutinases.

Substitution near N-terminal

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The variant of the invention has one or more amino acid substitutions in the vicinity of the N-terminal. The substitution is within a distance of 17 Å (e.g. within 12 Å) and/or within 20 positions (e.g. within 15 positions) of the N-terminal. The distance from the N-terminal is to be calculated between the Cα atom of the amino acids, and is calculated from an amino acid in a crystal structure (i.e. visible in the X-ray structure).

In the cutinase of *H. insolens* strain DSM 1800, the two N-terminal amino acids (Q1 and L2. i.e. Gln and Leu at positions 1 and 2) are not visible in the X-ray structure, so the distance is to be calculated from amino acid G3. Amino acids within 17 Å include positions 3-12, 18, 20-60, 62-64, 82, 85-86, 100-108, 110-111, 130-132, 174, 176-182, 184-185, 188, and 192. Those within 12 Å include positions 3-8, 25 22-27, 30-47, 53-59, 102, 177, and 180-181.

In the cutinase of *F. solani pisi*, the N-terminal amino acid G17 is visible in the X-ray structure. Amino acids within 17 Å include positions 17-26, 34-75, 77-79, 101, 115, 117-119, 147, 191-197, 199-200, and 203. Those within 12 Å include positions 17-22, 38, 40, 45-58, 60, 65, and 70-72.

The variants of the invention have improved thermostability compared to the parent enzyme. The thermostability may be determined from the denaturation tem-

5

perature by DSC (differential scanning calorimetry), e.g. as described in an example, e.g. at pH 8.5 with a scan rate of 90 K/hr. The variants may have a denaturation temperature which is at least 5°C higher than the parent enzyme.

The total number of substitutions in the above regions is typically 1-10, e.g. 1-5 substitutions in the above regions. In addition, the cutinase variant of the invention may optionally include other modifications of the parent enzyme, typically 10 or fewer, e.g. 5 or fewer alterations (substitutions, deletions or insertions) outside of the above regions. Thus, the total amino acid sequence of the variant typically 1-20, e.g. 1-10 alterations compared to the parent cutinase.

10 Solvent accessible surface

One or more of the substitutions may be made at an exposed amino acid residue, i.e. an amino acid residue having a solvent accessible surface. This can be calculated by the "dssp" program (version October 1988) described in W. Kabsch and C. Sander, Biopolymers, 22 (1983) pp. 2577-2637.

In the cutinase of *H. insolens* strain DSM 1800, the following amino acids lie within 17 Å of G3 at the N-terminal and have a solvent accessible surface greater than 0: 3-12, 18, 26-33, 36-38, 40-45, 47-56, 59-60, 62-64, 82, 85-86, 104-105, 174, 176-179, 181-182, 192.

Specific substitutions

The substitution near the N-terminal may specifically be one that increases the electrical charge, i.e. a substitution of a negatively charged amino acid with a neutral or positively charged amino acid or substitution of a neutral amino acid with a positively charged amino acid. Thus, a negative amino acid residue at a position corresponding to position E6, E10, E30, E47 D63, E82 and/or E179 in the cutinase of Humicola insolens strain DSM 1800 may be substituted by a neutral or positive amino acid, e.g. R, K, Y, H, Q or N. Some specific substitutions are those corresponding to E6Q/N, E10Q/N, E47K/R or E179Q/N. Also, a neutral amino acid residue at a position corresponding to N7, S11, N44 or N52 in the H. insolens cutinase may be substituted by a positive amino acid (R, K or H).

6

Another example of a substitution near the N-terminal is substitution with a Pro residue, e.g. a substitution corresponding to A14P or R51P in the cutinase of *Humicola insolens* strain DSM 1800.

Specific variants

The following are some examples of variants in the *H. insolens* cutinase. Corresponding variants may be made on the basis of other parent cutinases.

R51P

E6N/Q+ L138I

A14P+ E47K

10 E47K

E179N/Q

E6N/Q+ E47K+ R51P

A14P+ E47K+ E179N/Q

E47K+ E179N/Q

15 E47K+ D63N

E6N/Q+ E10N/Q+ A14P+ E47K+ R51P+ E179N/Q

E6N/Q+ A14P+ E47K+ R51P+ E179N/Q

Q1P+ L2V+ S11C+ N15T+ F24Y+ L46I+ E47K

Use of cutinase variant

The cutinase variant of the invention may be used, e.g., for the enzymatic hydrolysis of cyclic oligomers of poly(ethylene terephthalate), such as cyclic tri(ethylene terephthalate), abbreviated as c3ET.

In particular, this may be used to remove such cyclic oligomers from polyester containing fabric or yarn by treating the fabric or yarn with the cutinase variant, optionally followed by rinsing the fabric or yarn with an aqueous solution having a pH in the range of from about pH 7 to about pH 11. The treatment of polyester is conveniently carried out above the glass transition temperature of c3ET (about 55°C) and below the glass transition temperature of polyester (about 70°C). Thus, the treatment may suitably be carried out at 50-80°C, e.g. at 60-75°C. The process may be carried out in analogy with WO 97/27237.

7

The cutinase variant may be used to treat polyester-containing textile. e.g. PET (polymer of ethyleneglycol and terephthalic acid), P3GT (polymer of 1,3-propanediol and terephthalic acid) or a polyester/cotton blend. The treatment may provide benefits to the polyester textile such as improved wear and comfort, increased water permeability, reduced antistatic behavior, improve handle and softness, changed redeposition characteristics and/or color clarification.

The cutinase variant may be used to improve the functional finish of a PET-containing yarn or fabric by a treatment with the cutinase variant, followed by a treatment with a finishing agent such as a softener, an anti-crease resin, an anti-static agent, an anti-soiling agent or agents to impair wrinkle-free, permanent press ior fire resistance effects. The treatment with the cutinase variant may increase the number of functional groups in the surface, and this can be used to attach the functional finish. Examples of finishing agents are described in "SENSHOKU SIAGEKAKO BENRAN" published 1998-10-15 by Nihon Seni Sentaa KK.

The cutinase variant of the invention is also useful in detergents, where it may be incorporated to improve the removal of fatty soiling, as described in WO 94/03578 and WO 94/14964. The addition of the cutinase variant to laundry detergent may reduce malodor from cloth which is accumulated during several wash/wear-cycles.

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The cutinase variant may also be used for degradation and recycling of polyes-20 ter such as polycaprolactone (PCL), poly-ethyleneglycol-terephthalate (PET), polylactic acid, polybutylenesuccinate, and poly(hydroxybutiric acid)-co-(hydroxyvaleric acid), e.g. film and bottles, e.g. as described in JP-A 5-344897.

The cutinase variant may also be used for other known applications of lipases and cutinases, for example, in the baking industry (e.g. as described in WO 94/04035 and EP 585988), in the papermaking industry (e.g. for pitch removal, see EP 374700), and in the leather, wool and related industries (e.g. for degreasing of animal hides, sheepskin or wool), and for other applications involving degreasing/defatting. It may be used in immobilized form in the fat and oil industry, as a catalyst in organic synthesis (e.g. esterification, transesterification or ester hydrolysis reactions).

8

Dyeing polyester

The invention provides a process for dyeing polyester fabric or yarn. In this process, the fabric or yarn is first treated with a cutinase, e.g. 12-48 hours at 50-70°C or 65-70°C, pH 7-10, followed by dyeing with dye, e.g. a reactive dye, a disperse dye or a cationic dye. The reactive dye may be one that reacts with OH or COOH groups, e.g. having the structure Chromophore-NHPh-SO₂CH₂CH₂OSO₃Na. The dyeing may be conducted at 40-80°C, e.g. for 20-60 minutes.

The cutinase may be a thermostable cutinase having a thermal denaturation temperature, T_d, at pH 8.5 which is at least 5° higher than the parent cutinase, e.g. 7-10° higher, e.g. a value of 65°C or higher. The measurement may be made by DSC as described in an Example of this specification.

Surfactant

In the treatment of fabric or yarn, a conventional wetting agent and/or a dispersing agent may be used to improve the contact with the enzyme. The wetting agent may be a nonionic surfactant, e.g. an ethoxylated fatty alcohol. A very useful wetting agent is an ethoxylated and propoxylated fatty acid ester such as Berol 087 (product of Akzo Nobel, Sweden).

The dispersing agent may suitably be selected from nonionic, anionic, cationic, ampholytic or zwitterionic surfactants. More specifically, the dispersing agent may be selected from carboxymethylcellulose, hydroxypropylcellulose, alkyl aryl sulfonates, long-chain alcohol sulfates (primary and secondary alkyl sulfates), sulfonated olefins, sulfated monoglycerides, sulfated ethers, sulfosuccinates, sulfonated methyl ethers, alkane sulfonates, phosphate esters, alkyl isothionates, acylsarcosides, alkyltaurides, fluorosurfactants, fatty alcohol and alkylphenol condensates, fatty acid condensates, condensates of ethylene oxide with an amine, condensates of ethylene oxide with an amide, sucrose esters, sorbitan esters, alkyloamides, fatty amine oxides, ethoxylated monoamines, ethoxylated diamines, alcohol ethoxylate and mixtures thereof. A very useful dispersing agent is an alcohol ethoxylate such as Berol 08 (product of Akzo Nobel, Sweden).

9

Methods for preparing cutinase variants

The cutinase variant of the invention can be prepared by methods known in the art, e.g. as described in WO 94/14963 or WO 94/14964 (Unilever). The following describes methods for the cloning of cutinase-encoding DNA sequences, followed by methods for generating mutations at specific sites within the cutinase-encoding sequence.

Cloning a DNA sequence encoding a cutinase

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The DNA sequence encoding a parent cutinase may be isolated from any cell or microorganism producing the cutinase in question, using various methods well known in the art. First, a genomic DNA and/or cDNA library should be constructed using chromosomal DNA or messenger RNA from the organism that produces the cutinase to be studied. Then, if the amino acid sequence of the cutinase is known, labeled oligonucleotide probes may be synthesized and used to identify cutinase-encoding clones from a genomic library prepared from the organism in question. Alternatively, a labeled oligonucleotide probe containing sequences homologous to another known cutinase gene could be used as a probe to identify cutinase-encoding clones, using hybridization and washing conditions of lower stringency.

Yet another method for identifying cutinase-encoding clones would involve inserting fragments of genomic DNA into an expression vector, such as a plasmid, transforming cutinase-negative bacteria with the resulting genomic DNA library, and then plating the transformed bacteria onto agar containing a substrate for cutinase (i.e. maltose), thereby allowing clones expressing the cutinase to be identified.

Alternatively, the DNA sequence encoding the enzyme may be prepared synthetically by established standard methods, e.g. the phosphoroamidite method described S.L. Beaucage and M.H. Caruthers, (1981), Tetrahedron Letters 22, p. 1859-1869, or the method described by Matthes et al., (1984), EMBO J. 3, p. 801-805. In the phosphoroamidite method, oligonucleotides are synthesized, e.g. in an automatic DNA synthesizer, purified, annealed, ligated and cloned in appropriate vectors.

Finally, the DNA sequence may be of mixed genomic and synthetic origin, mixed synthetic and cDNA origin or mixed genomic and cDNA origin, prepared by

10

ligating fragments of synthetic, genomic or cDNA origin (as appropriate, the fragments corresponding to various parts of the entire DNA sequence), accordance with standard techniques. The DNA sequence may also be prepared by polymerase chain reaction (PCR) using specific primers, for instance as described in 5 US 4,683,202 or R.K. Saiki et al., (1988), Science 239, 1988, pp. 487-491.

Site-directed mutagenesis

Once a cutinase-encoding DNA sequence has been isolated, and desirable sites for mutation identified, mutations may be introduced using synthetic oligonucleotides. These oligonucleotides contain nucleotide sequences flanking the desired 10 mutation sites. In a specific method, a single-stranded gap of DNA, the cutinaseencoding sequence, is created in a vector carrying the cutinase gene. Then the synthetic nucleotide, bearing the desired mutation, is annealed to a homologous portion of the single-stranded DNA. The remaining gap is then filled in with DNA polymerase I (Klenow fragment) and the construct is ligated using T4 ligase. A specific example 15 of this method is described in Morinaga et al., (1984), Biotechnology 2, p. 646-639. US 4,760,025 discloses the introduction of oligonucleotides encoding multiple mutations by performing minor alterations of the cassette. However, an even greater variety of mutations can be introduced at any one time by the Morinaga method, because a multitude of oligonucleotides, of various lengths, can be introduced.

Another method for introducing mutations into cutinase-encoding DNA sequences is described in Nelson and Long, (1989), Analytical Biochemistry 180, p. 147-151. It involves the 3-step generation of a PCR fragment containing the desired mutation introduced by using a chemically synthesized DNA strand as one of the primers in the PCR reactions. From the PCR-generated fragment, a DNA fragment 25 carrying the mutation may be isolated by cleavage with restriction endonucleases and reinserted into an expression plasmid.

Expression of cutinase variants

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According to the invention, a DNA sequence encoding the variant produced by methods described above, or by any alternative methods known in the art, can be 30 expressed, in enzyme form, using an expression vector which typically includes con-

11

trol sequences encoding a promoter, operator, ribosome binding site. translation initiation signal, and, optionally, a repressor gene or various activator genes.

Expression vector

The recombinant expression vector carrying the DNA sequence encoding a cutinase variant of the invention may be any vector which may conveniently be subjected to recombinant DNA procedures, and the choice of vector will often depend on the host cell into which it is to be introduced. The vector may be one which, when introduced into a host cell, is integrated into the host cell genome and replicated to-10 gether with the chromosome(s) into which it has been integrated. Examples of suitable expression vectors include pMT838.

Promoter

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In the vector, the DNA sequence should be operably connected to a suitable promoter sequence. The promoter may be any DNA sequence which shows tran-15 scriptional activity in the host cell of choice and may be derived from genes encoding proteins either homologous or heterologous to the host cell.

Examples of suitable promoters for directing the transcription of the DNA sequence encoding a cutinase variant of the invention, especially in a bacterial host, are the promoter of the lac operon of E.coli, the Streptomyces coelicolor agarase 20 gene dagA promoters, the promoters of the Bacillus licheniformis α-amylase gene (amyL), the promoters of the Bacillus stearothermophilus maltogenic amylase gene (amyM), the promoters of the Bacillus amyloliquefaciens α -amylase (amyQ), the promoters of the Bacillus subtilis xylA and xylB genes etc. For transcription in a fungal host, examples of useful promoters are those derived from the gene encoding A. 25 oryzae TAKA amylase, the TPI (triose phosphate isomerase) promoter from S. cerevisiae (Alber et al. (1982), J. Mol. Appl. Genet 1, p. 419-434, Rhizomucor miehei aspartic proteinase, A. niger neutral α -amylase, A. niger acid stable α -amylase, A. niger glucoamylase, Rhizomucor miehei lipase, A. oryzae alkaline protease, A. oryzae triose phosphate isomerase or A. nidulans acetamidase.

Expression vector

The expression vector of the invention may also comprise a suitable transcription terminator and, in eukaryotes, polyadenylation sequences operably connected to the DNA sequence encoding the α -amylase variant of the invention. Ter-5 mination and polyadenylation sequences may suitably be derived from the same sources as the promoter.

The vector may further comprise a DNA sequence enabling the vector to replicate in the host cell in question. Examples of such sequences are the origins of replication of plasmids pUC19, pACYC177, pUB110, pE194, pAMB1 and pIJ702.

The vector may also comprise a selectable marker, e.g. a gene the product of which complements a defect in the host cell, such as the dal genes from B. subtilis or B. licheniformis, or one which confers antibiotic resistance such as ampicillin, kanamycin, chloramphenicol or tetracyclin resistance. Furthermore, the vector may comprise Aspergillus selection markers such as amdS, argB, niaD and sC, a marker 15 giving rise to hygromycin resistance, or the selection may be accomplished by cotransformation, e.g. as described in WO 91/17243.

The procedures used to ligate the DNA construct of the invention encoding a cutinase variant, the promoter, terminator and other elements, respectively, and to insert them into suitable vectors containing the information necessary for replication, 20 are well known to persons skilled in the art (cf., for instance, Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor, 1989).

Host Cells

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The cell of the invention, either comprising a DNA construct or an expression vector of the invention as defined above, is advantageously used as a host cell in 25 the recombinant production of a cutinase variant of the invention. The cell may be transformed with the DNA construct of the invention encoding the variant, conveniently by integrating the DNA construct (in one or more copies) in the host chromosome. This integration is generally considered to be an advantage as the DNA sequence is more likely to be stably maintained in the cell. Integration of the DNA con-30 structs into the host chromosome may be performed according to conventional

13

methods, e.g. by homologous or heterologous recombination. Alternatively, the cell may be transformed with an expression vector as described above in connection with the different types of host cells.

The cell of the invention may be a cell of a higher organism such as a mam-5 mal or an insect, but is preferably a microbial cell, e.g. a bacterial or a fungal (including yeast) cell.

Examples of suitable bacteria are Gram positive bacteria such as *Bacillus subtilis*, *Bacillus licheniformis*, *Bacillus lentus*, *Bacillus brevis*, *Bacillus stearothermophilus*, *Bacillus alkalophilus*, *Bacillus amyloliquefaciens*, *Bacillus coagulans*, *Bacillus coagulans*, *Bacillus circulans*, *Bacillus lautus*, *Bacillus megaterium*, *Bacillus thuringiensis*, or *Streptomyces lividans* or *Streptomyces murinus*, or gramnegative bacteria such as *E.coli*. The transformation of the bacteria may, for instance, be effected by protoplast transformation or by using competent cells in a manner known *per se*.

The yeast organism may favorably be selected from a species of *Saccharo-*15 myces or *Schizosaccharomyces*, e.g. *Saccharomyces* cerevisiae.

The host cell may also be a filamentous fungus e.g. a strain belonging to a species of Aspergillus, most preferably Aspergillus oryzae or Aspergillus niger, or a strain of Fusarium, such as a strain of Fusarium oxysporium, Fusarium graminearum (in the perfect state named Gribberella zeae, previously Sphaeria zeae, synonym with Gibberella roseum and Gibberella roseum f. sp. cerealis), or Fusarium sulphureum (in the prefect state named Gibberella puricaris, synonym with Fusarium trichothecioides, Fusarium bactridioides, Fusarium sambucium, Fusarium roseum, and Fusarium roseum var. graminearum), Fusarium cerealis (synonym with Fusarium crokkwellnse), or Fusarium venenatum.

In a preferred embodiment of the invention the host cell is a protease deficient or protease minus strain.

This may for instance be the protease deficient strain *Aspergillus oryzae* JaL 125 having the alkaline protease gene named "alp" deleted. This strain is described in WO 97/35956 (Novo Nordisk).

Filamentous fungi cells may be transformed by a process involving protoplast formation and transformation of the protoplasts followed by regeneration of the cell wall in a manner known per se. The use of *Aspergillus* as a host micro-organism

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is described in EP 238 023 (Novo Nordisk A/S), the contents of which are hereby incorporated by reference.

Production of cutinase variant by cultivation of transformant

The invention relates, *inter alia*, to a method of producing a cutinase variant of the invention, which method comprises cultivating a host cell under conditions conducive to the production of the variant and recovering the variant from the cells and/or culture medium.

The medium used to cultivate the cells may be any conventional medium suitable for growing the host cell in question and obtaining expression of the cutinase variant of the invention. Suitable media are available from commercial suppliers or may be prepared according to published recipes (e.g. as described in catalogues of the American Type Culture Collection).

The cutinase variant secreted from the host cells may conveniently be recovered from the culture medium by well-known procedures, including separating the cells from the medium by centrifugation or filtration, and precipitating proteinaceous components of the medium by means of a salt such as ammonium sulphate, followed by the use of chromatographic procedures such as ion exchange chromatography, affinity chromatography, or the like.

Expression of variant in plants

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The present invention also relates to a transgenic plant, plant part or plant cell which has been transformed with a DNA sequence encoding the variant of the invention so as to express and produce this enzyme in recoverable quantities. The enzyme may be recovered from the plant or plant part. Alternatively, the plant or plant part containing the recombinant enzyme may be used as such.

The transgenic plant can be dicotyledonous or monocotyledonous, for short a dicot or a monocot. Examples of monocot plants are grasses, such as meadow grass (blue grass, Poa), forage grass such as festuca, lolium, temperate grass, such as Agrostis, and cereals, e.g. wheat, oats, rye, barley, rice, sorghum and maize (corn).

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Examples of dicot plants are tobacco, legumes, such as lupins, potato, sugar beet, pea, bean and soybean, and cruciferous (family Brassicaceae), such as cauliflower, oil seed rape and the closely related model organism Arabidopsis thaliana.

Examples of plant parts are stem, callus, leaves, root, fruits, seeds, and tubers. In the present context, also specific plant tissues, such as chloroplast, apoplast, mitochondria, vacuole, peroxisomes and cytoplasm are considered to be a plant part. Furthermore, any plant cell, whatever the tissue origin, is considered to be a plant part.

Also included within the scope of the invention are the progeny of such plants, plant parts and plant cells.

The transgenic plant or plant cell expressing the variant of the invention may be constructed in accordance with methods known in the art. In short the plant or plant cell is constructed by incorporating one or more expression constructs encoding the enzyme of the invention into the plant host genome and propagating the resulting modified plant or plant cell into a transgenic plant or plant cell.

Conveniently, the expression construct is a DNA construct which comprises a gene encoding the enzyme of the invention in operable association with appropriate regulatory sequences required for expression of the gene in the plant or plant part of choice. Furthermore, the expression construct may comprise a selectable marker useful for identifying host cells into which the expression construct has been integrated and DNA sequences necessary for introduction of the construct into the plant in question (the latter depends on the DNA introduction method to be used).

The choice of regulatory sequences, such as promoter and terminator sequences and optionally signal or transit sequences is determined, eg on the basis of when, where and how the enzyme is desired to be expressed. For instance, the expression of the gene encoding the enzyme of the invention may be constitutive or inducible, or may be developmental, stage or tissue specific, and the gene product may be targeted to a specific tissue or plant part such as seeds or leaves. Regulatory sequences are eg described by Tague et al, Plant, Phys., 86, 506, 1988.

For constitutive expression the 35S-CaMV promoter may be used (Franck et al., 1980. Cell 21: 285-294). Organ-specific promoters may eg be a promoter from

16

storage sink tissues such as seeds, potato tubers, and fruits (Edwards & Coruzzi, 1990. Annu. Rev. Genet. 24: 275-303), or from metabolic sink tissues such as meristems (Ito et al., 1994. Plant Mol. Biol. 24: 863-878), a seed specific promoter such as the glutelin, prolamin, globulin or albumin promoter from rice (Wu et al., 5 Plant and Cell Physiology Vol. 39, No. 8 pp. 885-889 (1998)), a Vicia faba promoter from the legumin B4 and the unknown seed protein gene from Vicia faba described by Conrad U. et al, Journal of Plant Physiology Vol. 152, No. 6 pp. 708-711 (1998), a promotter from a seed oil body protein (Chen et al., Plant and cell physiology vol. 39, No. 9 pp. 935-941 (1998), the storage protein napA promoter from Brassica napus, 10 or any other seed specific promoter known in the art, eg as described in WO 91/14772. Furthermore, the promoter may be a leaf specific promoter such as the rbcs promoter from rice or tomato (Kyozuka et al., Plant Physiology Vol. 102, No. 3 pp. 991-1000 (1993), the chlorella virus adenine methyltransferase gene promoter (Mitra, A. and Higgins, DW, Plant Molecular Biology Vol. 26, No. 1 pp. 85-93 (1994), 15 or the aldP gene promoter from rice (Kagaya et al., Molecular and General Genetics Vol. 248, No. 6 pp. 668-674 (1995), or a wound inducible promoter such as the potato pin2 promoter (Xu et al, Plant Molecular Biology Vol. 22, No. 4 pp. 573-588 (1993).

A promoter enhancer element may be used to achieve higher expression of the enzyme in the plant. For instance, the promoter enhancer element may be an intron which is placed between the promoter and the nucleotide sequence encoding the enzyme. For instance, Xu et al. op cit disclose the use of the first intron of the rice actin 1 gene to enhance expression.

The selectable marker gene and any other parts of the expression construct may be chosen from those available in the art.

The DNA construct is incorporated into the plant genome according to conventional techniques known in the art, including *Agrobacterium*-mediated transformation, virus-mediated transformation, micro injection, particle bombardment, biolistic transformation, and electroporation (Gasser et al, Science, 244, 1293; Potrykus, Bio/Techn. 8, 535, 1990; Shimamoto et al, Nature, 338, 274, 1989).

Presently, Agrobacterium tumefaciens mediated gene transfer is the method of choice for generating transgenic dicots (for review Hooykas & Schilperoort, 1992.

17

Plant Mol. Biol. 19: 15-38), however it can also be used for transforming monocots, although other transformation methods are generally preferred for these plants. Presently, the method of choice for generating transgenic monocots is particle bombardment (microscopic gold or tungsten particles coated with the transforming DNA) of embryonic calli or developing embryos (Christou, 1992. Plant J. 2: 275-281; Shimamoto, 1994. Curr. Opin. Biotechnol. 5: 158-162; Vasil et al., 1992. Bio/Technology 10: 667-674). An alternative method for transformation of monocots is based on protoplast transformation as described by Omirulleh S, et al., Plant Molecular biology Vol. 21, No. 3 pp. 415-428 (1993).

Following transformation, the transformants having incorporated the expression construct are selected and regenerated into whole plants according to methods well-known in the art.

MATERIALS AND METHODS

Plasmids

15 pJSO026

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This is a *S. cerevisiae* expression plasmid described in WO 97/07205 and in J.S.Okkels, (1996) "A URA3-promoter deletion in a pYES vector increases the expression level of a fungal lipase in Saccharomyces cerevisiae. Recombinant DNA Biotechnology III: The Integration of Biological and Engineering Sciences, vol. 782 of the Annals of the New York Academy of Sciences).

pFuku83

This is a yeast and E. coli shuttle vector for expression of the H. insolens cutinase under the control of a TPI promoter, constructed from pJSO026.

Substrate

25 BETEB

Terephthalic acid bis(2-hydroxyethyl)ester dibenzoate is herein abbreviated as BETEB (benzoyl-ethylene-terephthalic-ethelene-benzoate). It was prepared from terephthalic acid bis (2-hydroxyethyl) ester and benzoic acid.

Lipase activity (LU)

A substrate for lipase is prepared by emulsifying tributyrin (glycerin tributyrate) using gum Arabic as emulsifier. The hydrolysis of tributyrin at 30 °C at pH 7 is followed in a pH-stat titration experiment. One unit of lipase activity (1 LU) equals the amount of enzyme capable of releasing 1 µmol butyric acid/min at the standard conditions.

Differential scanning calorimetry (DSC)

Sample and reference solutions are carefully degassed immediately prior to loading of samples into the calorimeter (reference: buffer without enzyme). Sample and reference solutions (approx. 0.5 ml) are thermally pre-equillibrated for 20 minutes at 5°C. The DSC scan is performed from 5 C to 95 C at a scan rate of approx. 90 K/hr. Denaturation temperatures are determined at an accuracy of approx. +/- 1 C. A VP-DSC from MicroCal Inc. is suitable for the experiments.

Methods

15 PCR conditions

step 1: 94° C, 120 sec.

step 2: 94° C, 60 sec

step 3: 50° C, 60 sec

step 4: 72° C, 150 sec.

Go to step 2, 35 cycles

step 5: 72° C, 480 sec.

Step 6: 4° C, for ever

EXAMPLES

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Example 1: Preparation of cutinase variants

A DNA sequence encoding *H. insolens* cutinase was obtained as described in US 5,827,719 (Novo Nordisk) and was found to have the DNA sequence shown in SEQ ID NO: 1 therein.

19

Variants were prepared by localized random mutagenesis and selection of positive clones by incubation at 60°C for 1 day on BETEB plates. The BETEB plates contained 200 ml/l of 500 mM glycine buffer (pH 8.5), 1.25 g/l of BETEB (dissolved in hot ethanol) and 20 g/l of agar.

Three positive variants were isolated, and their amino acid sequence was determined. They were found to have the following modifications, compared to the parent *H. insolens* cutinase:

A14P + E47K

E47K

10 E179Q

Example 2: Site directed mutation

A variant of the *H.* insolens cutinase having the substitutions E6Q+ E47K+ R51P was prepared as follows:

A pair of PCR primers were designed so as to introduce amino acid substitu-15 tions, making use of the existed restriction enzyme sites nearby, as follows (an asterisk indicates an introduced mutation):

Upper primer: E6Q F

cgg cag ctg gga gcc atc c*ag aac

Pvu II

20 Lower primer: E47K,R51P

cgc cct gga tcc aga tgt tcg* gga tgt ggg act t*aa ggc

BamH I

PCR was run using these primers and pFukuNL83 as a template under the PCR condition described above.

The obtained PCR fragment was purified by Clontech Spincolumn and digested with *Pvu* II and *BamH* I.

The resultant fragment was gel-purified and ligated to pFukuNL83 which had been digested with the same restriction enzyme sites.

Example 3: Thermostability of cutinase variants

Variants

The thermostability was tested as described below for the *H. insolens* cutinase and the following variants thereof:

5 A14P+ E47K

E47K

E179Q

E6Q+ E47K+ R51P

A14P+ E47K+ E179Q

10 E6Q+ A14P+ E47K+ R51P+ E179Q

E6Q+ E10Q+ A14P+ E47K+ R51P+ E179Q

Differential Scanning Calorimetry (DSC)

Thermostability of cutinase variants was investigated by means of DSC at pH 4.5 (50 mM acetate buffer) and pH 8.5 (50 mM glycyl-glycine buffer). The thermal denaturation temperature, T_d, was taken as the top of denaturation peak (major endothermic peak) in thermograms (Cp vs. T) obtained after heating of enzyme solutions at a constant programmed heating rate.

The parent cutinase was found to have T_d of 63°C at pH 8.5. Six of the above variants were found to have T_d of 70-73°C, i.e. an improvement of 7-10°.

The parent cutinase was found to have T_d of 61°C at pH 4.5. Five of the above variants were found to have T_d of 64-66°C, i.e. an improvement of 3-5°.

Hydrolysis of BETEB

The thermostability of the *H. insolens* cutinase and two of the above variants was measured by hydrolysis of BETEB at elevated temperature. For each cutinase, the following mixture was incubated for 17 hours at various temperatures in the range 55-70°C:

- 0.1 ml 0.5 M glycyl-glycine buffer (pH 8.5)
- 0.1 ml 0.5 % BETEB dissolved in ethanol
- 0.1 ml enzyme solution (approx. 25 LU/ml)
- 30 0.7 ml Milli Q water

The degree of hydrolysis was measured after the incubation. The results are shown in the table below.

·	Variant	Variant	Parent
	27 LU/ml	25 LU/ml	24 LU/ml
55°C	98 %	99 %	72 %
60°C	91 %	83 %	33 %
65°C	66 %	13 %	7 %

These results clearly show that the variants have improved thermostability compared to the parent cutinase.

Hydrolysis of BETEB

The thermostability of the *H. insolens* cutinase and three of the above variants was measured by hydrolysis of BETEB at 60°C for 2 hours. The hydrolysis was carried out at the above conditions, except that the temperature was fixed at 60°C and the cutinase dosage was varied. The results below are shown in the table below.

LU/ml	Variant	Variant	Variant	Parent
0	0 %	0 %	0 %	0 %
10	97 %	99 %	9 %	6 %
20	98 %	99 %	74 %	
50	98 %	94 %	93 %	15 %
100	88 %	69 %	92 %	34 %
300				41 %
600				63 %
1200				82 %

The results show a much faster hydrolysis at 60°C with the variants than with the parent cutinase.

22

Example 4: Hydrolysis of c3ET

The *H. insolens* cutinase and five of the above variants were tested in hydrolysis of c3ET at elevated temperature. For each cutinase, the following mixture was incubated for 2 hours at various temperatures.

5 0.115mg c3ET (0.1ml of 2mM c3ET dissolved in HFIP was taken in reaction vessel. Solvent was removed under vacuum, then dried up at 70°C over night)

0.1ml 0.5M glycyl-glycine buffer (pH8.5)

0.1ml enzyme solution (approx. 600LU/ml)

0.8ml Milli Q water

After the incubation, 2ml of 1,1,1,3,3,3-Hexafluoro-2-propanol (HFIP) was added to each reaction mixture, then hydrolysis ratio was measured by HPLC. The results shown in Fig 3 clearly indicate that the variants have improved thermostability compared to the parent cutinase.

Example 5: Hydrolysis of c3ET on varn

The thermostability of the *H. insolens* cutinase five of the above variants was tested using polyester yarn containing c3ET as by product. The following substrate mixture was preincubated at 60 or 65°C:

0.1g polyester yarn

0.2ml 0.5M glycyl-glycine buffer (pH8.5)

20 1.7ml Milli Q water

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After preincubation, 0.1ml enzyme solution (approx. 1000 LU/ml) was added to each reaction vessel and incubated for 17 hours. Then 2ml HFIP was added and left for 30 minutes to extract and hydrolyze c3ET sitting on the surface of the polyester yarn; then the hydrolysis ratio was measured. The results are shown in Fig. 4.

It is seen that the variants are more effective than the parent cutinase for hydrolyzing c3ET on polyester yarn. One variant gives higher hydrolysis ratio at 65°C than at 60°C.

Example 6: Treatment of yarn with cutinase variant

Time courses of c3ET hydrolysis on polyester yarn at different temperature or dosage were examined. Time course at different temperatures is shown in Fig 5. It is seen that the optimum temperature is 65°C. At 70°C there is still about half of the activity left. Time course with increased enzyme dosage is shown in Fig 6. The curves at dosage 275 and 550 LU/ml are seen to be the same, indicating that the hydrolysis ratio reached to plateau between dosage of 100 to 275 LU/ml. Presumably 200LU/ml is enough.

Example 7: Dyeing polyester with reactive dye

The following polyester fabrics were treated:

woven fabric; ca. 2 x 2 cm, 34mg

knitted fabric; ca. 1.5 x 1.5 cm, 50mg

Each fabric was soaked in 0.9 ml, 50 mM GlyGly (glycyl-glycine) buffer (pH 8.5) and 0.1 ml solution of a variant of the *H. insolens* cutinase (1100 LU/ml), and incubated at 65 or 70°C. After one day, another 0.1 ml enzyme solution was added, incubation was continued for two more days, the fabrics were then taken out and rinsed in water. A comparative experiment was made with the parent cutinase, and a blank was treated in the same manner without enzyme.

The fabrics were stirred in a mixture of 9 g 120 g Na₂SO₄ and 60 g Na₂CO₃ in 3 liter deionized water at 60 °C for 30 min, and then rinsed with running warm water. The reactive dye was Celmazol Brilliant Blue B (product of Mitsui Chemical Co., Japan), which has the structure Chromophore-NHPh-SO₂CH₂CH₂OSO₃Na.

In all four experiments, (woven and knitted, 65 and 70°C), the fabrics were uniformly dyed.

25 Example 8: Solubilization of polyester fragments from knitted textile

A 1x1 cm sample of knitted polyester textile (PET, polymer of ethyleneglycol and terephthalic acid) was incubated for 1 hour in 1 ml of buffer at pH 10, 60°C with 0.01 mg of a variant of *H. insolens* cutinase. The reaction mixture was separated, and the release of terephthalic acid was found by measuring OD at 250 nm (ex-

PCT/DK99/00678 WO 00/34450

24

pressed as OD₂₅₀/mg PET) comparative experiments made without were enzyme or with the parent cutinase. Results:

	Enzyme	OD ₂₅₀
Invention	Cutinase variant	4.5
Reference	Parent cutinase	0.3
	None	0.1

The results show that the variant is effective in solubilizing polyester.

In another experiment, the cutinase variant was tested for 2 hours at 65°C 5 with and without the addition of a non-ionic surfactant (alcohol ethoxylate, product name Softanol 50), using various amounts of the variant from 0.5 to 200 LU/ml. The results showed more solubilization in the presence of non-ionic surfactant.

Example 9: Hydrolysis of polycaprolactone and polyester film

About 0.1 g of polycaprolactone or polyester film were put in tubes. They were soaked in 5ml of 50mM GlyGly buffer (pH 8.5) with or without a variant of H. insolens cutinase (450 LU). They were incubated at 70°C for 5 hours. After the reaction we observed a thin layer of hydrolysate on the surface of the tubes with enzyme, both with polycaprolactone and with polyester film. On the other hand no change 15 was observed in controls without enzyme. In the case of polycaprolactone there was 10% of weight loss. We see no weight change of polyester.

Example 10: cPET hydrolysis

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The performance of a cutinase variant was compared with the parent enzyme (H. insolens cutinase). The trials were done as follows:

An oligomer-stained swatch of (black) PET-fabric (app. 4cm x 13cm) is subjected to the enzyme-treatment at relatively low agitation in a so-called minitergitometer apparatus. The PET-fabric is mounted onto a cylindrical, perforated holder (radius ca.2 cm, height ca 6 cm), that rotates around its axis, and with the oligomer stained side of the PET fabric facing the exterior of the cylinder.

The fabric is immersed in a 150ml glass-beaker containing 100ml of the treatment solution at a given temperature (here 65°C). After a given treatment time

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(here 90minutes) the PET swatch is removed from the bath and rinsed in deionized water and air dried.

After conditioning the swatches are visually ranked (with respect to oligomer stain removal) on the side having the oligomer-staining. The rating being as follows:

-2: Sample significantly worse than blank (no enzyme)

-1: Sample slightly worse than blank (no enzyme)

0: Sample can not be distinguished from blank

1: Sample slightly improved vs blank

2: Sample significantly improved over blank

Also, the swatches are read spectrofotometrically (apparatus: Hunterlab Reflectometer) to quantify the color strength (K/S-value at 600nm).

The table below summarizes the test-conditions for a trial comparing the performance the enzymes under similar conditions:

Temperature:

65°C

Buffer/pH:

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50 mM glycine buffer, pH 10.3

Treatment time (min)

90

Dosage of Enzyme (LU/g)

30000

15 Results from the trial are summarized below

Enzyme	Visual rating (avg.)	K/S Difference @ 600 nm
None	0 (defined)	2.33
Parent cutinase	0	2.38
Cutinase variant	1.5-2.0	2.89

From this set of experiments it thus appears that the parent enzyme provides no or only very limited effect at the given test conditions (probably because the temperature is too high for the enzyme to retain activity), while the cutinase variant provides a substantial removal of the oligomer staining from the PET-fabric.

Example 11: cPET hydrolysis

The pH and temperature profile of a variant of *H. insolens* cutinase was tested in a model disperse dyeing experiment. The trials were performed as follows:

An oligomer-stained swatch of (black) PET-fabric is subjected to the conditions of a typical disperse dyeing sequence in a Werner Mathis Labornat. In overview of the process, the swatch is added to a buffer solution, heated to 130°C,
cooled down to the treatment temperature. Enzyme or buffer is added and then held
at the desired temperature for 30 minutes. The solution is cooled down to room temperature and turbidity in the wash liquor is measured. The reduction in turbidity is a
direct measure of the cutinase activity, corresponding to hydrolyzed cPET oligomers.

Detailed description of the experiment:

A black PET (app. 4cm \times 13cm) swatch is added 140 ml 100 mM Britton-Robinson buffer containing 0.2 g/l Lutensol AT11 (BASF) and loaded in the Laborat (32 rotation per minute).

The Laborat is heated to 130°C at a gradient of 9°C/minute, and held for 10 minutes.

The beakers are cooled to run temperature (according to table below) at a gradient of 9°C/minute, and held for 1 minute.

10 mL enzyme solution (100 LU/ml) of the variant) or buffer solution (0 LU/ml) 20 at appropriate pH is injected to the beakers.

The Labornat is re-heated to temperature at a gradient of 2°C/minute, and held for 30 minutes.

The swatches are removed, and the wash liquor is cooled down to room temperature.

Turbidity of the wash liquors are measured.

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Evaluation: Turbidity is measured on Hach 18900 Ratio Turbidimeter (standardized with 1.8, 18, and 180 NTU Turbidity Standards). Enzyme performance is calculated relative to a blank as the difference between turbidity of blank liquor (no enzyme) and turbidity of enzyme treated liquor.

The relative performance (reduction in turbidity) of the cutinase variant is calculated, and the results are shown in the following table. When a negative num-

27 "negative". A negative number is ber is obtained, then the result is given as assumed to be an artifact, caused by the variation of the set up.

Temperature	pH 7	pH 8	pH 9	pH 10
60°C	39	57	37	14
65°C	39	16	60	30
70°C	25	12	54	33
75°C	22	50	114	58
85°C	negative	negative	15	negative

The results show that the cutinase variant is active over a broad pH and 5 temperature range, with optimum oligomer removal in the current set up around pH 9 and 75°C. Inactivation seems to occur at or above 85°C.

Example 12: cPET hydrolysis

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The effect of treatment time was investigated for a variant of H. insolens cutinase in a model disperse dyeing experiment. The trials were performed as follows:

An oligomer-stained swatch of (black) PET-fabric is subjected to the conditions of a typical disperse dyeing sequence in a Werner Mathis Labomat. In overview of the process, the swatch is added to a buffer solution, heated to 130°C, cooled down to the treatment temperature. Enzyme or buffer (100 mM Britton-Robinson pH 9) is added, and then held at 75°C for 0-40 minutes. The solution is 15 cooled down to room temperature and turbidity in the wash liquor is measured. The reduction in turbidity is a direct measure of the cutinase activity, corresponding to hydrolyzed cPET oligomers.

Detailed description of the experiment:

A black PET (app. 4cm x 13cm) swatch is added to 140 ml 100 mM Britton-20 Robinson buffer containing 0.2 g/l Lutensol AT11 (BASF) and loaded in the Laborat (32 rotation per minute).

The Laborat is heated to 130°C at a gradient of 9°C/minute, and the temperature is held for 10 minutes.

The beakers are cooled to 75°C at a gradient of 9°C/minute, and held for 1 25 minute.

10 mL enzyme solution (100 LU/ml of variant) or 100 mM Britton-Robinson buffer pH 9.0 (0 LU/ml) is injected into the beakers.

The Laborat is re-heated to 75°C at a gradient of 2°C/minute, and held for the appropriate number of minutes (0-40 minutes, see table below).

The swatches are removed, and the wash liquor is cooled down to room temperature.

Turbidity of the wash liquors are measured.

Evaluation: Turbidity is measured on Hach 18900 Ratio Turbidimeter (standardized with 1.8, 18, and 180 NTU Turbidity Standards). Enzyme performance is calculated relative to a blank at time equal to zero: Turbidity of blank liquor at time zero (no enzyme) subtracted turbidity of enzyme treated liquor (at a given time).

The relative performance (reduction in turbidity) of the cutinase variant was calculated, and the results are shown in the following table.

Time (minutes)	Relative perform- ance (Reduction in turbidity)
0	0
5	42
10	48
15	62
20	69
25	85
30	72
40	78

The results show that the effect of the enzyme is increased over time. At the current enzyme dose and oligomer concentration, it seems to level off above approx. 20 minutes.

Example 13: Fiber modification

The effect on wetting characteristics of a disperse dyed polyester fabric was investigated by treating the fabric with a variant of *H. insolens* cutinase prior to dye-

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ing. The experiment therefore consisted of two phases, the actual fiber modification and the disperse dyeing pro- cedure.

Phase 1 - Fiber Modification:

Equipment:

Atlas Launder-O-meter LP2

Fabric:

knit 100 % scoured polyester from Testfabrics

:Ha

50 mM potassium phosphate buffer, pH 7

Abrasives:

5 big steel balls

Beaker Vol.:

120 mL

Treatment:

2 hours 65°C then ramped up to 90°C and held for 1 hour

Swatch Prep:

Cut 3* 1.5 g swatch of fabric, 3 per beaker = 4.5 g

Rinse:

5 Rinse in deionized water.

Phase 2 - Dyeing - disperse dye:

Dye Solution:

Add together with deionized water to make liquor ratio 1:20-

0.4 % Dianix Red (DyStar) SE-CB (owf)

10 pH to 4.5 - 5

Dyeing Procedure:

- 1. One swatch per treatment from the fiber modification is used for the dyeing (1.5 g/swatch is used for the liquor ratio calculation).
- Make dyebath according to the recipe above. Add the cold dye solution
 to the Laborat beakers and heat to 55°C at a gradient of 3.5°C/minute. Run for 5 minutes once temperature has been reached.
 - 3. Add the fabric to the beaker.
 - 4. Raise temperature to 130°C at a gradient of 1.5°C/minute. Dye for 30 minutes.
- 5. Cool to 70°C at a gradient of 5°C/minute. Drop bath, but collect, and rinse fabric hot (60°C) for 10 minutes. Follow the hot rinse with a room temperature overflow rinse until all bleeding had stopped.
 - 6. Let air dry overnight.

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Tests/Analysis:

AATCC Test Method 61 - Colorfastness to washing

Percent Dyebath Exhaustion - Spectrophotometer

K/S and L* - Reflectometer

5 AATCC TM-79 Drop Test

Results:

The results from the fiber modification are shown in the following table.

Variant dosage	Staining (AATCC TM- 61)	Color Change (K/S @ 530 before and after TM-61)	Drop Test (AATCC TM-79)
Blank	4.5	5	53 sec.
50 LU/mL	4.5	5	18 sec.
100 LU/mL	4.5	5	15 sec.

The results show that the treatment of polyester with the variant increases the wetting substantially. No adverse effects are noticed on the dyeability with the disperse dye in the current set-up.

Example 14: Malodor reduction in textiles soiled with human sweat/sebum by use of a cutinase variant in laundry

The performance of cutinase, with respect to malodor reduction, can be tested in a one-cycle washing trial carried out in a Terg-O-tometer.

Experimental conditions:

Washing liquor: 1000 ml per beaker

Swatches: 100 % polyester (interlock knitted, previously cleaned by Soxhlet extraction). 24 swatches (3.3 × 3.5 cm) per beaker.

Soil: Human male axillary sweat and sebum applied by wiping the armpits after exercise.

Detergent: 5 g/L of a standard color detergent. No pH adjustment.

Water hardness: 3.2 mM Ca²⁺/Mg²⁺ (in a ratio of 5:1)

Wash Temperature: 30°C

25 Wash time: 30 min

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Rinse: 15 minutes in running tap water

Evaluation:

After wash the wet swatches are placed in closed, tinted 200 ml glasses. A trained sensory panel (9-11 judges) evaluates the odor by sniffing the headspace over the wet samples and evaluates the total odor intensity. The odor intensity is noted by placing a mark on an unstructured line scale measuring 15 cm, with word anchors at each end ('nothing' at the beginning of the scale and 'very strong' at the end). All evaluations are performed twice. The swatches are evaluated on day 1, 2 and 3 after wash (swatches are kept in the glasses at all times).

CLAIMS

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- 1. A variant of a parent fungal cutinase, which variant:
 - a) comprises substitution of one or more amino acid residues at a position which is located:
 - i) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 20 positions from the N-terminal amino acid, and
 - b) is more thermostable than the parent cutinase.
- 10 2. The variant of the preceding claim which comprises substitution of one or more amino acid residues at a position which is located:
 - within 12 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 15 positions from the N-terminal amino acid.
 - 3. A variant of a parent fungal cutinase comprising substitution of one or more amino acid residues which is located:
 - a) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - b) within 20 positions from the N-terminal amino acid,

with the proviso that it is not a variant of the cutinase of *Fusarium solani pisi* having one of the substitutions R17, T18, T19V, D21N, I24E, Y38F, R40, G41A, S42, T43, E44, T45, G46, N47R, G49, T50, L51, P53, S54, A56C, S57, N58R, S61, A62E, K65A, D66S, G67D, W69Y, I70C, G74, G75, R78, Y119, G192, P193, D194R, 25 A195, R196, G197V, or A199C (*Fusarium solani pisi* cutinase numbering).

- 4. A variant of a parent fungal cutinase comprising substitution of one or more amino acid residues which:
 - a) has a solvent accessible surface, and

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- b) is located:
 - i) within 17 Å from the location of the Nterminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 20 positions from the N-terminal amino acid,

with the proviso that it is not a variant of the cutinase of *Fusarium solani pisi* having one of the substitutions T18, Y38F, R40, G41A, S42, T43, E44, T45, N47R, G49, T50, L51, P53, S54, A56C, A62E or G192 (*Fusarium solani pisi* cutinase numbering).

- 10 5. A variant of a parent fungal cutinase comprising substitution of one or more amino acid residues which is located:
 - a) less than 12 Å from the location of the N-terminal amino group (as calculated from amino acid residues in a crystal structure), and/or
 - b) within 15 positions from the N-terminal amino acid,
- with the proviso that the variant is not the cutinase of *Fusarium solani pisi* having one of the substitutions R17, T18, T19V, D21N, Y38F, R40, T45, G46, N47R, G49, T50, L51, P53, S54, A56C, S57, N58R, K65A or I70C (*Fusarium solani pisi* cutinase numbering).
- 6. The variant of any preceding claim wherein the parent cutinase is native to a filamentous fungus, preferably a strain of *Humicola* or *Fusarium*, preferably *H. insolens* or *F. solani pisi*, most preferably *H. insolens* strain DSM 1800.
 - 7. The variant of any preceding claim wherein the parent cutinase has an amino acid sequence which can be aligned with the cutinase of *H. insolens* strain DSM 1800.
- 25 8. The variant of any preceding claim wherein the parent cutinase has an amino acid sequence which is at least 50 % homologous to the cutinase of *H. insolens* strain DSM 1800, preferably at least 70 % homologous, more preferably at least 80 % homologous.

- 9. A variant of a parent fungal cutinase from *Humicola insolens* which comprises substitution of one or more amino acid residues located:
 - a) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
- b) within 20 positions from the N-terminal amino acid.

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- 10. The variant of the preceding claim which comprises substitution of one or more amino acid residues located:
 - a) less than 12 Å from the location of the N-terminal amino group (as calculated from amino acid residues in a crystal structure), and/or
 - b) within 15 positions from the N-terminal amino acid
- 11. The variant of any preceding claim which comprises substitution of one or more amino acids having a solvent accessible surface.
- 12. The variant of any preceding claim wherein one or more substitutions is substitution of a negatively charged amino acid with a neutral or positively charged amino acid or substitution of a neutral amino acid with a positively charged amino acid.
- 13. The variant of the preceding claim wherein one or more substitutions is at a position corresponding to position E6, E10, E30, E47, D63, E82 and/or E179 in the cutinase of *Humicola insolens* strain DSM 1800, preferably a substitution with 20 R/K/Y/H/Q/N, more preferably a substitution corresponding to E6N/Q, E10N/Q, E47K/R and/or E179N/Q (*H. insolens* cutinase numbering).
 - 14. The variant of any preceding claim wherein one or more substitutions is substitution with a Pro residue, preferably at a position corresponding to position A14 and/or R51.
- 25 15. The variant of any preceding claim which has one, two, three, four, five or six of said substitutions.

- 16. The variant of any preceding claim which has substitutions corresponding to one of the following in the cutinase of *Humicola insolens* strain DSM 1800:
 - a) R51P
 - b) E6N/Q + L138I
- 5 c) A14P + E47K
 - d) E47K
 - e) E179N/Q
 - f) E6N/Q + E47K + R51P
 - g) A14P + E47K + E179N/Q
- 10 h) E47K + E179N/Q
 - i) E47K + D63N
 - i) E6N/Q + A14P + E47K + R51P + E179N/Q
 - k) E6N/Q + E10N/Q + A14P + E47K + R51P + E179N/Q, or
 - l) Q1P + L2V + S11C + N15T + F24Y + L46I + E47K
- 15 17. The variant of any preceding claim which has hydrolytic activity towards terephthalic acid esters, particularly towards cyclic tri(ethylene terephthalate) and/or Terephthalic acid bis(2-hydroxyethyl)ester dibenzoate (BETEB).
 - 18. The variant of any preceding claim which has a denaturation temperature which is at least 5° higher than the parent cutinase, preferably measured at pH 8.5
- 20 19. A DNA sequence encoding the variant of any preceding claim.
 - 20. A vector comprising the DNA sequence of the preceding claim.
 - 21. A transformed host cell harboring the DNA sequence of claim 19 or the vector of claim 20.
 - 22. A method of producing the variant of any of claims 1-18 comprising
- 25 a) cultivating the cell of claim 21 so as to express and preferably secrete the variant, and

WO 00/34450 PCT/DK99/00678

36

b) recovering the variant.

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- 23. A method of constructing a cutinase variant, which method comprises:
 - a) selecting a parent fungal cutinase,
- 5 b) identifying one or more amino acid residues in the parent cutinase at positions which are:
 - i) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 20 positions from the N-terminal amino acid, and
 - c) making alterations each of which is an insertion, a deletion or a substitution of the amino acid residue,
 - d) optionally, making alterations each of which is an insertion, a deletion or a substitution of an amino acid residue at one or more positions other than b),
 - e) preparing the variant resulting from steps b-d,
 - f) testing the thermostability of the variant,
 - g) optionally repeating steps b-f, and
- h) selecting a variant having higher thermostability than the parent cuti-20 nase.
 - A method of producing a cutinase variant, which method comprises:
 - a) selecting a parent fungal cutinase,
 - b) identifying one or more amino acid residues in the parent cutinase at positions which are:
 - i) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 20 positions from the N-terminal amino acid, and
- c) making alterations each of which is an insertion, a deletion or a substitution of the amino acid residue,

WO 00/34450

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d) optionally, making alterations each of which is an insertion, a deletion or a substitution of an amino acid residue at one or more positions other than b),

37

PCT/DK99/00678

- e) preparing the variant resulting from steps b-d,
- f) testing the thermostability of the variant,
- g) optionally repeating steps b-f,
- h) selecting a variant having higher thermostability than the parent cutinase, and
- i) producing the variant to obtain the cutinase variant.
- 10 25. A process for enzymatic hydrolysis of a cyclic oligomer of poly(ethylene terephthalate), which process comprises treating the cyclic oligomer with a variant of a parent fungal cutinase, which variant comprises substitution of one or more amino acid residues at a position which is located:
 - i) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 20 positions from the N-terminal amino acid.
 - 26. The process of the preceding claim, in which the cyclic oligomer is cyclic tri(ethylene terephthalate).
- 20 27. The process of claim 25 or 26 wherein the treatment is done at 60-80°C, preferably at 65-75°C.
 - 28. The process of any of claims 25-27 wherein the cyclic oligomer is present in and on the fibers of a polyester containing fabric or yarn.
- 29. The process of any of claims 25-28 which further comprises subsequently rinsing the fabric or yarn, preferably rinsing with an aqueous solution having a pH in the range of from about pH 7 to about pH 11.
 - 30. A process for dyeing polyester fabric or yarn, comprising:

WO 00/34450 PCT/DK99/00678

38

a) treating the fabric or yarn with a cutinase having a thermal denaturation temperature of 65°C or higher at pH 8.5; and

- b) dyeing the treated fabric with a reactive dye or a disperse dye.
- 5 31. The process of the preceding claim wherein the cutinase is the variant of any of claims 1-18.
 - 32. A detergent composition comprising a surfactant and the variant of any of claims 1-18.
- 33. A method for detecting cutinase activity in a sample, comprising incubating the sample with terephthalic acid bis(2-hydroxyethyl)ester dibenzoate and detecting hydrolysis of said ester.
 - 34. A process for improving the functional finish of a PET-containing yarn or fabric comprising
 - a) treating the yarn or fabric with the variant of any of claims 1-18, and
- 15 b) subsequently the yarn or fabric with a finishing agent selected from the group consisting of softeners, anti-crease resins, anti-static agents, anti-soiling agents.

WO 00/34450 PCT/DK99/00678

Fig. 1
3D structure of cutinase from *Humicola insolens*

	-		GT 37	70	_	04 404	7 025	10 200	1 00 46 72
ATOM	1	N	GLY		3	24.424		18.390	1.00 46.73
ATOM	2	CA	GLY		3	23.848		17.546	1.00 42.29
ATOM	3	С	GLY		3		-10.112	16.727	1.00 37.35
MOTA	4	0	GLY	Α	3	25.347		16.728	1.00 35.38
ATOM	5	N	ALA	Α	4	23.664	-10.625	15.797	1.00 34.53
ATOM	6	CA	ALA	Α	4	23.051	-10.874	14.555	1.00 30.95
ATOM	7	С	ALA	Α	4	21.574	-11.246	14.920	1.00 28.33
ATOM	8	0	ALA		4	20.677		14.446	1.00 22.94
ATOM	9	СВ	ALA		4	23.574	-11.780	13.556	1.00 26.92
ATOM	10	N		A	5	21.583	-12.058	16.043	1.00 26.48
ATOM	11	CA		Α	5	20.281	-12.289	16.637	1.00 25.65
ATOM	12	C		A	5	20.316		18.118	1.00 23.03
	13	0		A	5				
ATOM					5	21.060		18.717	1.00 24.74
ATOM	14	CB	ILE	A			-13.683	16.524	1.00 26.04
ATOM	15	CG1		A	5	19.852	-13.927	15.050	1.00 29.85
ATOM	16	CG2		A	5	18.374		17.159	1.00 20.48
ATOM	17	CD1		Α	5		-15.133	14.709	1.00 27.96
ATOM	18	И	GLU	A	6	19.461	-11.377	18.668	1.00 20.52
ATOM	19	CA	GLU	Α	6	19.207	-11.015	20.040	1.00 17.94
ATOM	20	С	GLU	Α	6	17.711	-11.027	20.432	1.00 17.76
MOTA	21	0	GLU	Α	6	16.931	-10.165	19.990	1.00 17.60
ATOM	22	CB	GLU	A	6	19.809	-9.614	20.199	1.00 14.22
ATOM	23	CG	GLU	Α	6	21.232	-9.374	20.385	1.00 16.71
ATOM	24	CD	GLU	A	6	22.148	-10.387	21.030	1.00 34.47
ATOM	25	OE1	GLU		6	21.634	-11.347	21.693	1.00 49.57
ATOM	26	OE2	GLU		6	23.410	-10.310	20.975	1.00 37.43
ATOM	27	N	ASN		7	17.375	-11.895	21.333	1.00 21.67
ATOM	28	CA	ASN		7	16.070	-11.854	21.846	1.00 24.04
ATOM	29	C	ASN		7	15.927	-11.488	23.238	1.00 22.08
ATOM	30	Õ	ASN		7	15.098	-12.179	23.820	1.00 24.00
ATOM	31	CB	ASN		7	15.468	-13.307	21.820	1.00 25.06
		CG	ASN		7				
ATOM	32					15.039	-13.160	20.341	1.00 38.52
ATOM	33	OD1	ASN		7	15.519	-14.147	19.759	1.00 48.45
ATOM	34	ND2	ASN		7	14.318	-12.081	19.968	1.00 36.89
MOTA	35	N	GLY		8	16.671	-10.813	23.926	1.00 23.56
ATOM	36	CA	GLY		8		-10.628	25.363	1.00 23.69
ATOM	37	С	GLY	A	8	15.366	-10.247	25.984	1.00 22.72
ATOM	38	0	GLY	Α	8	14.967	-10.939	26.867	1.00 32.25
ATOM	39	N	LEU	Α	9	14.785	-9.144	25.755	1.00 23.61
ATOM	40	CA	LEU	Α	9	13.470	-8.753	26.033	1.00 23.73
ATOM	41	С	LEU	Α	9	12.559	-9.961	25.782	1.00 25.93
ATOM	42	0	LEU	Α	9		-10.054	26.480	1.00 30.47
ATOM	43	СВ	LEU		9	12.971	-7.621	25.105	1.00 5.84
ATOM	44	CG	LEU		9	11.556		25.470	1.00 23.25
ATOM	45		LEU		9	11.422	-6.765	26.968	1.00 20.21
ATOM	46		LEU		9		-6.071	24.714	1.00 17.64
ATOM	47	N	GLU		10		-10.786	24.713	1.00 17.04
111 011	- '	74	010	17	10	12.113	-10.100	22.113	1.00 29.36

7.004	40	CA	GLU	70.	10	11 635	-11.681	24.484	1.00	33.93
ATOM	48		GLU		10		-12.872	25.412		32.18
ATOM	49	С			10		-13.159	25.996		36.67
ATOM	50	0	GLU					23.012		40.97
MOTA	51	CB	GLU		10		-12.303	22.745		51.96
ATOM	52	CG	GLU		10			21.437		54.08
ATOM	53	CD	GLU		10		-11.711	20.635		48.22
MOTA	54	OE1	GLU		10		-11.440			52.39
ATOM	55	OE2	GLU		10		-11.643	21.471		29.58
ATOM	56	N	SER		11		-13.334	25.688		
ATOM	57	CA	SER		11		-14.455	26.645		35.25
ATOM	58	C	SER		11		-14.012	28.047		39.86
ATOM	59	0	SER		11		-14.790	28.919		43.72
ATOM	60	CB	SER		11.		-15.364	25.983		33.73
ATOM	61	OG	SER	A	11		-14.620	25.928		46.98
ATOM	62	N	\mathtt{GLY}	Α	12 .	13.467	-12.802	28.456		41.40
ATOM	63	CA	\mathtt{GLY}	Α	12	13.841	-12.332	29.752		45.34
ATOM	64	С	\mathtt{GLY}	Α	12		-12.562	30.694		47.62
ATOM	65	0	GLY	Α	12		-12.335	30.335	1.00	50.76
ATOM	66	N	SER	Α	13	12.969	-12.900	31.936		48.09
ATOM	67	CA	SER	Α	13	11.974	-13.158	32.995		45.26
ATOM	68	С	SER	Α	13		-11.933	33.772		39.53
ATOM	69	0	SER	Α	13	12.563	-11.204	33.992		36.30
ATOM	70	CB	SER	A	13	12.708	-14.006	34.101		51.20
ATOM	71	OG	SER	Α	13	12.006	-13.947	35.338		57.14
ATOM	72	N	ALA	A	14	10.256	-11.785	34.214		35.22
ATOM	73	CA	ALA	Α	14	10.068	-10.530	34.964	1.00	34.78
ATOM	74	С	ALA	Α	14	10.574	-10.620	36.417	1.00	37.51
ATOM	75	0	ALA	Α	14	10.809	-9.584	37.113	1.00	38.41
ATOM	76	СВ	ALA	Α	14	8.714	-9.915	34.903	1.00	32.71
ATOM	77	N	ASN	Α	15	11.039	-11.834	36.737	1.00	38.85
ATOM	78	CA	ASN	A	15	11.715	-12.086	37.963	1.00	43.49
ATOM	79	C	ASN		15	13.073	-11.411	37.953	1.00	46.45
ATOM	80	Ö	ASN		15	13.453	-11.022	39.022	1.00	52.50
ATOM	81	СВ	ASN		15	12.088	-13.533	38.207	1.00	53.08
ATOM	82	CG	ASN		15	10.772	-14.226	38.553	1.00	71.86
ATOM	83		ASN		15	9.837	-13.535	38.998	1.00	71.73
ATOM	84		ASN		15	10.866	-15.523	38.267	1.00	77.71
ATOM	85	N	ALA		16		-11.305	36.812	1.00	46.73
ATOM	86	CA	ALA		16		-10.470	36.743	1.00	41.22
ATOM	87	C	ALA		16	15.031		35.798	1.00	36.70
ATOM	88	Ö	ALA		16	16.027		35.075	1.00	37.67
ATOM	89	СВ	ALA		16		-11.545	36.301		41.80
	90	N	CYS		17	14.300		35.843		30.62
ATOM	91	CA	CYS		17	14.614		34.997		31.78
ATOM	92	C	CYS		17	16.024		35.149		32.94
ATOM	93	0	CYS		17	16.744		36.113		39.10
ATOM		CB	CYS		17	13.679		35.138		28.00
ATOM	94		CYS		17	12.048		34.858		24.72
ATOM	95	SG	CIS	, A.	1	12.040	0.505	54.000		

ATOM	96	N	PRO		18	16.529	-5.910	34.092		30.49
ATOM	97	CA	PRO		18	17.994	-5.626	33.971		22.04
ATOM	98	С	PRO	Α	18	18.178	-4.138	34.241	1.00	20.15
ATOM	99	0	PRO	Α	18	17.085	-3.459	34.370	1.00	17.83
ATOM	100	CB	PRO	Α	18	18.353	-6.003	32.559	1.00	19.20
ATOM	101	CG	PRO	Α	18	17.044	-6.595	32.101	1.00	20.16
ATOM	102	CD	PRO	Α	18	15.903	-5.936	32.792	1.00	24.35
ATOM	103	N	ASP	Α	19	19.428	-3.652	34.011	1.00	14.85
ATOM	104	CA	ASP	A	19	19.451	-2.168	34.226	1.00	
ATOM	105	С		A	19	18.739	-1.367	33.156	1.00	
ATOM	106	0	ASP		19	18.311	-0.242	33.430		23.84
ATOM	107	СВ		A	19	20.896	-1.818	34.485	1.00	
ATOM	108	CG	ASP		19	21.433	-2.389	35.793		42.30
ATOM	109	OD1	ASP		19	21.162	-3.549	36.297	1.00	
ATOM	110	OD2	ASP		19	22.251	-1.719	36.543		54.02
ATOM	111	N	ALA		20	18.646	-1.780	31.895	1.00	
ATOM	112	CA	ALA		20	18.066	-1.036	30.809	1.00	17.43
ATOM	113	C	ALA		20	17.713	-2.087	29.703	1.00	16.06
ATOM	114	0	ALA		20	18.334	-3.172	29.860	1.00	
ATOM	115	CB	ALA		20	18.975	-0.048	30.100	1.00	9.45 12.07
ATOM	116	N	ILE		21	16.814				
	117	CA	ILE				-1.602	28.829	1.00	8.47
ATOM					21	16.657	-2.583	27.753	1.00	9.23
ATOM	118	C	ILE		21	16.952	-1.745	26.486	1.00	14.77
ATOM	119	0	ILE		21	16.681	-0.473	26.403	1.00	12.01
ATOM	120	CB		A	21	15.208	-2.984	27.837	1.00	16.28
ATOM	121	CG1		A	21	14.851	-3.898	28.956	1.00	15.55
ATOM	122	CG2		A	21	14.689	-3.671	26.514		13.71
ATOM	123	CD1		A	21	13.401	-3.879	29.372	1.00	6.12
ATOM	124	N	LEU		22	17.432	-2.451	25.391		12.24
ATOM	125	CA	LEU		22	17.665	-1.774	24.087		11.27
MOTA	126	С	LEU		22	16.849	-2.517	23.038	1.00	14.60
ATOM	127	0	LEU		22	16.908	-3.781	22.850	1.00	9.78
ATOM	128	CB	LEU		22	19.087	-1.865	23.693	1.00	10.96
ATOM	129	CG	LEU	Α	22	19.493	-1.543	22.257	1.00	10.32
ATOM	130	CD1	LEU	Α	22	19.311	-0.081	21.900	1.00	4.72
ATOM	131	CD2	LEU	Α	22	20.990	-1.842	22.156	1.00	7.42
ATOM	132	N	ILE	Α	23	16.038	-1.815	22.242	1.00	15.13
ATOM	133	CA	ILE	A	23	15.298	-2.459	21.115		18.06
ATOM	134	С	ILE		23	15.916	-1.771	19.901		17.42
ATOM	135	0	ILE		23	16.117	-0.519	19.795		19.31
ATOM	136	СВ	ILE		23	13.820	-2.194	21.392		18.16
ATOM	137	CG1			23	13.208	-3.076	22.447		14.23
ATOM	138	CG2	ILE		23	12.787	-2.167	20.247		13.19
ATOM	139	CD1	ILE		23	12.142	-2.065	22.976		20.41
ATOM	140	N	PHE		24	16.218	-2.548	18.940		14.59
ATOM	141	CA	PHE		24	16.859	-2.159	17.671		11.72
ATOM	142	C	PHE		24	16.347	-2.719	16.353	1.00	
ATOM	143	0	PHE		24	16.095	-3.998	16.161		
ALON	147	0	LUC	n	44	10.033	-3.990	10.101	1.00	3.47

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ATOM	144	CB	PHE		24	18.195	-2.855	17.658		12.61
ATOM	145	CG	PHE		24	19.015	-2.150	16.716	1.00	10.72
ATOM	146	CD1	PHE	Α	24	19.457	-0.844	16.913	1.00	13.08
ATOM	147	CD2	PHE	Α	24	19.325	-2.852	15.558	1.00	6.61
ATOM	148	CE1	PHE	Α	24	20.232	-0.187	15.983	1.00	4.86
ATOM	149	CE2	PHE		24	20.061	-2.218	14.545	1.00	7.61
ATOM	150	CZ	PHE		24	20.550	-0.823	14.804	1.00	8.78
ATOM	151	N	ALA		25	16.037	-1.700	15.449	1.00	6.32
ATOM	152	CA	ALA		25	15.662	-2.158	14.068	1.00	7.18
ATOM	153	C	ALA		25	16.851	-1.976	13.055	1.00	8.59
ATOM	154	0	ALA		25	17.518	-1.000	13.133	1.00	5.95
ATOM	155	CB	ALA		25	14.488	-1.402	13.562	1.00	8.27
ATOM	156	N	ARG		26	17.174	-3.032	12.325	1.00	8.84
ATOM	157	CA	ARG	Α	26	18.134	-3.278	11.277	1.00	4.04
ATOM	158	С	ARG	Α	26	17.691	-2.694	9.894	1.00	7.67
ATOM	159	0	ARG	Α	26	16.527	-2.361	9.525	1.00	9.36
ATOM	160	CB	ARG	Α	26	18.581	-4.659	10.756	1.00	6.06
ATOM	161	CG	ARG		26	17.705	-5.741	10.439	1.00	5.08
ATOM	162	CD	ARG		26	18.069	-7.224	10.382	1.00	6.73
ATOM	163	NE	ARG		26	17.000	-8.053	9.708	1.00	9.04
ATOM	164	CZ	ARG		26	15.724	-8.206	9.912	1.00	7.06
ATOM	165	NH1	ARG		26	15.724		10.895		
							-7.535			22.93
ATOM	166	NH2	ARG		26	14.809	-8.825	9.346	1.00	7.89
ATOM	167	N	GLY		27	18.761	-2.539	9.092	1.00	7.71
ATOM	168	CA	GLY		27	18.537	-1.888	7.782	1.00	5.34
ATOM	169	С	GLY		27	18.063	-2.896	6.862	1.00	4.70
ATOM	170	0	GLY	A	27	18.155	-4.139	7.075	1.00	13.14
MOTA	171	N	SER	Α	28	17.562	-2.612	5.765	1.00	11.82
ATOM	172	CA	SER	A	28	17.108	-3.325	4.615	1.00	14.72
ATOM	173	С	SER	A	28	18.214	-4.327	4.142	1.00	7.74
ATOM	174	0	SER	Α	28	19.286	-3.973	4.083	1.00	6.71
ATOM	175	CB	SER		28	16.460	-2.352	3.538	1.00	6.38
ATOM	176	OG	SER		28	16.819	-0.978	3.833		28.10
ATOM	177	N	THR		29	17.942	-5.634	4.241	1.00	4.79
ATOM	178	CA	THR		29					
		CA				18.562	-6.763	3.914	1.00	8.71
ATOM	179		THR		29	19.500	-7.271	4.985		14.00
ATOM	180	0	THR		29	20.162	-8.326	4.713		17.68
ATOM	181	CB	THR		29	19.454	-6.680	2.617		14.90
ATOM	182	OG1	THR		29	20.736	-6.066	2.595	1.00	14.00
ATOM	183	CG2	THR	A	29	18.785	-5.888	1.561	1.00	15.59
ATOM	184	N	GLU	Α	30	19.740	-6.599	6.105	1.00	14.52
ATOM	185	CA	GLU	A	30	20.677	-7.266	7.056	1.00	14.10
ATOM	186	С	GLU		30	20.092	-8.513	7.647		13.07
ATOM	187	0	GLU		30	18.916	-8.726	7.705	1.00	19.98
ATOM	188	СВ	GLU		30	21.228	-6.371	8.072	1.00	15.45
ATOM	189	CG	GLU		30	21.166	-4.945	7.709	1.00	8.37
ATOM	190	CD	GLU		30	22.073				23.08
							-4.143	8.637		
ATOM	191	OE 1	GLU	A	30	21.395	-3.328	9.284	1.00	19.26

PCT/DK99/00678

ATOM	192 ⁻	OE2	GLU	A	30	23	.317	-4.327	8.712	1.00	19.71
ATOM	193	N	PRO		31	20	.875	-9.479	7.918	1.00	13.09
ATOM	194	CA	PRO		31			-10.818	8.402	1.00	14.56
ATOM	195	С	PRO		31			-10.698	9.895	1.00	18.27
ATOM	196	0	PRO		31	20	.148	-9.636	10.392	1.00	20.45
ATOM	197	CB	PRO		31			-11.692	8.215	1.00	10.95
ATOM	198	CG	PRO		31			-10.664	8.455	1.00	11.24
ATOM	199	CD	PRO		31		.350	-9.316	7.864	1.00	13.71
ATOM	200	N	\mathtt{GLY}		32		-	-11.689	10.472	1.00	18.99
ATOM	201	CA	GLY		32			-11.774	11.816	1.00	13.53
ATOM	202	С	GLY		32		.133	-10.808	12.188	1.00	16.62
ATOM	203	0	GLY		32	•	.345	-10.294	11.411	1.00	17.01
ATOM	204	N	ASN		33		.055	-10.528	13.468	1.00	16.15
ATOM	205	CA	ASN		33		.290	-9.346	13.823	1.00	14.74
ATOM	206	С	ASN		33		.294	-8.273	14.230	1.00	15.46
ATOM	207	0	ASN		33		.774	-7.184	14.575	1.00	15.90
ATOM	208	CB	ASN		33		.241	-9.663	14.867	1.00	
ATOM	209	CG	ASN		33		.827	-10.201	16.127	1.00	
ATOM	210	OD1	ASN		33		.112	-10.395	17.089		19.05
ATOM	211	ND2	ASN		33		.074	-10.460	16.112	1.00	13.29
ATOM	212	N		A	34		.633	-8.378	14.282	1.00	14.22
ATOM	213	CA		A	34		.282	-7.171	14.751	1.00	12.97
ATOM	214	С		A	34		.142	-6.663	13.611	1.00	19.02
ATOM	215	0		A	34		.654	-5.512	13.713	1.00	26.04
ATOM	216	CB		A	34		.202	-7.329	15.859	1.00	13.39
ATOM	217	CG		A	34		.579	-7.713	17.163	1.00	9.02
ATOM	218	SD		A	34		.175	-6.316	18.069	1.00	9.13
ATOM	219	CE		A	34		.481	-5.121	18.095	1.00	4.11
ATOM	220	N	GLY		35		.259	-7.446	12.550	1.00	19.99
ATOM	221	CA	GLY		35		.071	-7.135	11.418	1.00	14.30
ATOM	222	С	GLY		35		.511	-7.340	11.764	1.00	17.58
ATOM	223	0	GLY		35		.965	-7.724	12.842	1.00	12.78
ATOM	224	N		A	36		.450	-6.839	10.950	1.00	20.63
ATOM	225	CA		Α	36		.833	-7.029	11.277	1.00	17.71
ATOM	226	C		A	36		.609	-5.714	11.280	1.00	16.15
ATOM	227	0		A	36		.865	-5.618	11.662	1.00	20.30
ATOM	228	CB	ILE		36		.412	-8.070	10.327		30.19
ATOM	229		ILE		36		.088	-7.448	8.959		31.16
ATOM	230		ILE		36		.944	-9.490	10.543		15.68
ATOM	231	CD1			36		.922	-8.149	7.958		34.10
ATOM	232	N	THR		37		.905	-4.589	11.040		13.00
ATOM	233	CA	THR		37		.825	-3.396	11.141		9.67
ATOM	234	С	THR		37		.587	-2.513	12.350		15.44
ATOM	235	0	THR		37		.040	-3.055	13.410		20.20
ATOM	236	CB	THR		37		.592	-2.679	9.818		14.13
ATOM	237	OG1			37		.241	-2.212	9.503	1.00	22.62
ATOM	238	CG2	THR		37		.949		8.800	1.00	
ATOM	239	N	VAL	A	38	25	.733	-1.493	12.249	1.00	11.92

3 500 6	0.40			_								
ATOM	240	CA	VAL		38		237	-0.80		.411		15.22
ATOM	241	C	VAL		38		588	-1.45		.612	1.00	
ATOM	242	0	VAL		38		906	-1.18		.733	1.00	
ATOM	243	CB	VAL		38		124	0.18		.855	1.00	
ATOM	244	CG1	VAL		38		663	0.89		.167	1.00	
ATOM	245	CG2	VAL		38		570	1.02		.670	1.00	6.75
ATOM	246	N	\mathtt{GLY}		39		745	-2.41	0 14	.677	1.00	
ATOM	247	CA	GLY	Α	39	23.	135	-3.15		.746	1.00	11.03
ATOM	248	С	GLY	Α	39	24.	096	-3.58	6 16	.791	1.00	13.34
ATOM	249	0	${ t GLY}$	Α	39	24.	131	-3.18	1 17	.934	1.00	15.13
ATOM	250	N	PRO	A	40	25.	067	-4.34	0 16	.352	1.00	14.70
ATOM	251	CA	PRO	Α	40	. 26.	094	-5.02	5 17	.171	1.00	13.44
ATOM	252	С	PRO	Α	40	27.		-3.90	9 17	.589		11.81
ATOM	253	0	PRO	Α	40	27.		-3.87		.764		12.79
ATOM	254	CB	PRO	Α	40	26.		-6.11		.279	1.00	8.43
ATOM	255	CG	PRO		40	25.		-6.24		.950	1.00	4.84
ATOM	256	CD	PRO		40	25.		-4.90		.995	1.00	12.36
MOTA	257	N	ALA		41	27.		-2.97		.695	1.00	7.41
ATOM	258	CA	ALA		41	28.		-1.96		.278		11.03
ATOM	259	C	ALA		41	27.		-1.20		.439		14.87
ATOM	260	Ö	ALA		41	28.		-0.50		.274	1.00	14.26
ATOM	261	CB	ALA		41	28.		-0.90		.313	1.00	7.17
ATOM	262	N	LEU		42	26.		-0.81		.237		11.87
ATOM	263	CA	LEU		42	25.		-0.04		.300		12.36
ATOM	264	C	LEU		42	25.				.624		
ATOM	265	0	LEU		42	25.		-0.85				11.94
ATOM	266	СВ	LEU					-0.39		.730	1.00	8.33
					42	24.		0.168		.811		13.24
ATOM	267	CG	LEU		42	23.:		1.160		.676	1.00	6.90
ATOM	268	CD1	LEU		42	24.		2.419		.962	1.00	6.62
ATOM	269	CD2	LEU		42	21.		1.580		.943	1.00	7.11
ATOM	270	N	ALA		43	24.		-2.09		.482		10.88
ATOM	271	CA	ALA		43	24.		-3.02		.553		12.37
ATOM	272	С	ALA		43	26.		-3.13		.252		15.45
ATOM	273	0	ALA		43	25.		-2.743		.433		20.80
ATOM	274	CB	ALA		43	24.		-4.32		.002	1.00	9.60
ATOM	275	N	ASN		44	27.	263	-3.440	21	.636	1.00	16.91
ATOM	276	CA	ASN	Α	44	28.	454	-3.43	4 22	.439	1.00	20.33
ATOM	277	С	ASN	A	44	28.	717	-2.04	4 23	.113	1.00	17.66
ATOM	278	0	ASN	Α	44	29.	019	-1.99	L 24	.301	1.00	17.06
ATOM	279	CB	ASN	Α	44	29.	756	-3.69	5 21	.625	1.00	35.48
ATOM	280	CG	ASN	Α	44	29.	564	-5.11	5 21	.138		58.23
ATOM	281	OD1	ASN	A	44	30.		-5.403		.034		79.77
ATOM	282	ND2	ASN	Α	44	28.		-5.945		.921		70.10
ATOM	283	N	GLY		45	28.		-0.988		.297		14.39
ATOM	284	CA	GLY		45	29.		0.22		.976		11.65
ATOM	285	С	GLY		45	28.		0.25		.234		14.30
ATOM	286	Ō	GLY		45	28.		0.582		.385		10.77
ATOM	287	N	LEU		46	26.		0.099		.065		16.88
					- 0	20.	U U I	0.09.	_ 27	. 0 0 0	1.00	10.00

ATOM		CA LEU		25.96	8 0.24	8 25.207	7 1 00 16 00
ATOM		C LEU		26.39			
ATOM	_	O LEU		26.57			
ATOM		CB LEU	A 46	24.608			
ATOM		CG LEU		23.642			
ATOM		CD1 LEU .	A 46	24.089			
ATOM		D2 LEU		22.275			
ATOM	295 N	GLU Z	A 47	26.523			
ATOM		A GLU	A 47	26.910			
ATOM	297 C	GLU 2	A 47	28.140			
ATOM	298 C	GLU A		28.722			
ATOM	299 C	B GLU A		27.147		-	
ATOM	300 C	G GLU A		27.386			1.00 33.33
ATOM	301 C			27.661		. – . •	1.00 51.29
ATOM	302 o	E1 GLU A		26.741			1.00 68.40
ATOM	303 0	E2 GLU A		28.856			1.00 66.37
ATOM	304 N			28.992		26.830	1.00 78.70
ATOM	305 C				-1.626	27.215	1.00 27.50
ATOM	306 C			30.331	-1.518	27.789	1.00 25.23
ATOM	307 o			30.108	-0.555	28.926	1.00 26.91
ATOM	308 CI			31.124	-0.058	29.462	1.00 33.39
ATOM	309 00			31.116	-0.990	26.621	1.00 21.90
ATOM	310 N	HIS A		31.294	0.422	26.483	1.00 27.87
ATOM	311 CA			28.826	-0.101	28.995	1.00 25.04
ATOM	312 C	HIS A		28.542	0.955	29.956	1.00 19.72
ATOM	313 0	HIS A	49	27.480	0.461	30.950	1.00 22.55
ATOM	314 CE		49	27.186	1.089	31.898	1.00 27.93
ATOM	315 CG		49	28.094	2.197	29.463	1.00 16.13
ATOM		1 HIS A	49	28.806	3.036	28.520	1.00 39.79
ATOM		2 HIS A	49	29.564	4.058	28.953	1.00 45.66
ATOM	318 CE		49	28.776	3.070	27.197	1.00 46.91
ATOM	319 NE		49	30.028	4.750	27.979	1.00 45.87
ATOM	320 N	ILE A	50	29.544	4.139	26.934	1.00 50.84
ATOM	321 CA		50	27.009	-0.703	30.715	1.00 18.34
ATOM	322 C	ILE A	50	25.874	-1.129	31.415	1.00 19.89
ATOM	323 0	ILE A		25.917	-2.629	31.146	1.00 26.29
ATOM	324 CB	ILE A	50	25.322	-3.023	30.168	1.00 25.33
ATOM	_	l ILE A	50	24.527	-0.535	31.008	1.00 10.50
ATOM		ILE A	50 50	24.340	0.906	31.292	1.00 4.97
ATOM	327 CD1		50	23.466	-1.298	31.697	1.00 12.96
ATOM	328 N	ARG A	50	23.413	1.845	30.602	1.00 16.65
ATOM	329 CA	ARG A	51	26.707	-3.256	32.066	1.00 31.77
ATOM	330 C	ARG A	51	26.887		32.107	1.00 29.06
ATOM	331 0	ARG A	51	25.457	-5.331	32.170	1.00 32.68
ATOM	332 N	ARG A	51 50	25.396	-6.363		1.00 37.16
ATOM	333 CA	ASN A	52	24.380	-4.817	32.788	1.00 28.48
ATOM	334 C	ASN A	52	23.284	-5.767	32.832	1.00 26.39
ATOM	335 0	ASN A	52		-5.178		1.00 27.75
	222 0	ASN A	52	21.333			1.00 26.68

ATOM	336	CB	ASN		52	22.750	-5.884	34.232	1.00 34.86
ATOM	337	CG	ASN		52	21.637	-6.879	34.271	1.00 39.54
ATOM	338		ASN		52	20.781	-6.541	35.095	1.00 54.31
ATOM	339	ND2	ASN		52	21.611	-7.954	33.503	1.00 48.82
ATOM	340	N	ILE	Α	53	22.127	-5.699	30.800	1.00 24.42
ATOM	341	CA	ILE	Α	53	21.261	-5.092	29.772	1.00 20.15
ATOM	342	С	ILE	Α	53	20.585	-6.151	28.912	1.00 17.63
ATOM	343	0	ILE	Α	53	21.020	-7.349	28.917	1.00 18.01
ATOM	344	CB	ILE	Α	53	22.245	-4.297	28.880	1.00 14.09
ATOM	345	CG1	ILE	Α	53	21.682	-3.257	27.936	1.00 22.91
ATOM	346	CG2	ILE	Α	53	22.907	-5.321	27.946	1.00 16.37
ATOM	347	CD1	ILE	A	53 .	22.877	-2.315	27.622	1.00 38.17
ATOM	348	N	TRP		54	19.447	-5.880	28.383	1.00 15.19
ATOM	349	CA	TRP		54	18.804	-6.889	27.567	1.00 17.96
ATOM	350	С	TRP		54	18.803	-6.230	26.151	1.00 19.82
ATOM	351	ō	TRP		54	18.340	-5.059	25.985	1.00 13.32
ATOM	352	CB	TRP		54	17.364	-7.046	27.998	1.00 23.18
ATOM	353	CG			54	16.949	-7.932	29.100	1.00 23.10
ATOM	354	CD1	TRP		54	17.757	-8.727	29.895	1.00 24.37
ATOM	355	CD2			54	15.595	-8.164	29.603	1.00 24.40
ATOM	356	NE1	TRP		54	17.004	-9.372	30.858	1.00 30.21
ATOM	357	CE2	TRP		54	15.692	-9.039	30.700	1.00 23.87
ATOM	358	CE3	TRP		54	14.358			
ATOM	359	CZ2	TRP		54		-7.633	29.243	1.00 36.26
ATOM	360	CZ3	TRP		54	14.611	-9.442	31.432	1.00 19.75
ATOM	361	CH2	TRP		54 54	13.316	-8.042	30.009	1.00 32.94
ATOM	362		ILE			13.451	-8.916	31.068	1.00 23.02
ATOM	363	N			55	19.063	-7.152	25.204	1.00 15.21
		CA	ILE		55	19.178	-6.655	23.838	1.00 12.41
ATOM	364	C	ILE		55	18.091	-7.215	22.962	1.00 11.40
ATOM	365	0	ILE		55	17.955	-8.378	22.680	1.00 7.34
ATOM	366	CB	ILE		55	20.546	-6.962	23.201	1.00 16.44
ATOM	367	CG1	ILE		55	21.939	-6.409	23.702	1.00 8.75
ATOM	368	CG2	ILE		55	20.384	-6.460	21.750	1.00 21.77
ATOM	369	CD1	ILE		55	21.767	-5.582	24.863	1.00 16.23
ATOM.	370	N	GLN		56	17.226	-6.412	22.390	1.00 9.67
ATOM	371	CA	GLN		56	16.161	-7.016	21.619	1.00 10.90
ATOM	372	С	GLN		56	16.432	-6.621	20.143	1.00 13.08
ATOM	373	0	GLN		56	16.402	-5.393	19.953	1.00 10.32
ATOM	374	CB	GLN		56	14.786	-6.542	22.014	1.00 11.49
ATOM	375	CG	GLN	Α	56	13.653	-7.256	21.316	1.00 23.47
ATOM	376	CD	GLN	Α	56	13.789	-8.741	21.351	1.00 24.88
ATOM	377	OE1	GLN	Α	56	13.610	-9.379	20.324	1.00 9.56
ATOM	378	NE2	GLN	Α	56	14.119	-9.221	22.544	1.00 17.94
ATOM	379	N	GLY	Α	57	16.288	-7.645	19.216	1.00 6.84
ATOM	380	CA	GLY	A	57	16.174	-7.019	17.841	1.00 16.15
ATOM	381	С	GLY	Α	57	14.740	-7.085	17.267	1.00 13.72
ATOM	382	0	GLY	Α	57	14.124	-8.016	17.752	1.00 12.70
ATOM	383	N	VAL		58	14.068	-6.264	16.525	1.00 12.73

ATOM	384	CA	VAL		58		2.739	-6.308	16.070		11.16
ATOM	385	С	VAL		58		2.715	-7.246	14.893		14.85
ATOM	386	0	VAL		58		3.234	-6.891	13.849		18.64
ATOM	387	CB	VAL		58		2.262	-4.984	15.352	1.00	6.54
ATOM	388	CG1			58		0.894	-4.974	14.731	1.00	5.89
ATOM	389	CG2	VAL	Α	58	13	2.650	-3.840	16.331	1.00	5.86
ATOM	390	N	GLY	Α	59	1.3	2.209	-8.465	15.008	1.00	21.96
ATOM	391	CA	GLY	Α	59	13	2.120	-9.385	13.874	1.00	17.81
ATOM	392	С	GLY	A	59	10	0.645	-9.561	13.550	1.00	23.35
ATOM	393	0	GLY	A	59	9	9.919	-8.579	13.249	1.00	27.99
ATOM	394	N	GLY	Α	60	1	0.166	-10.805	13.623	1.00	18.75
ATOM	395	CA	GLY	A	60	1	8.841	-11.142	13.285	1.00	11.46
ATOM	396	С	GLY	Α	60	;	3.550	-10.833	11.851	1.00	14.56
ATOM	397	0	GLY	Α	60	9	9.160	-11.439	11.003	1.00	16.32
ATOM	398	N	PRO	A	61			-10.103	11.612	1.00	12.10
ATOM	399	CA	PRO	Α	61		7.123	-9.774	10.250	1.00	14.70
ATOM	400	С	PRO		61		3.230	-8.941	9.570		22.17
ATOM	401	0	PRO		61		8.143	-8.758	8.344	1.00	
ATOM	402	СВ	PRO		61		5.911	-8.860	10.332	1.00	
ATOM	403	CG	PRO		61		5.880	-8.514	11.784	1.00	
ATOM	404	CD	PRO		61		6.723	-9.417	12.576	1.00	
ATOM	405	N	TYR		62		9.162	-8.257	10.292	1.00	
ATOM	406	CA	TYR		62		9.973	-7.242	9.674	1.00	
ATOM	407	C	TYR		62		1.133	-7.907	9.047	1.00	
ATOM	408	Ö	TYR		62		2.132	-8.213	9.691	1.00	
ATOM	409	CB	TYR		62		0.504	-6.401	10.803	1.00	
ATOM	410	CG	TYR		62		1.461	-5.421	10.236	1.00	
ATOM	411	CD1	TYR		62		1.343	-4.920	9.032	1.00	
ATOM	412	CD2	TYR		62		2.465	-4.971	10.969	1.00	
ATOM	413	CE1	TYR		62		2.206	-3.997	8.506	1.00	19.28
ATOM	414	CE2	TYR		62		3.438	-4.101	10.490	1.00	
ATOM	415	CZ	TYR		62		3.327	-3.571	9.186	1.00	
ATOM	416	OH	TYR		62		4.320	-2.649	8.791	1.00	
ATOM	417	N	ASP		63		0.998	-8.419	7.816	1.00	
ATOM	418	CA	ASP		63		2.137	-9.011	7.010	1.00	
ATOM	419	CA	ASP		63		3.027	-7.973	6.453		17.97
ATOM	420	0	ASP		63		3.628	-8.442	5.512		14.94
	421	_							6.015		17.16
ATOM	421	CB	ASP		63		1.474	-9.873			
ATOM		CG	ASP		63		0.563				27.75
ATOM	423		ASP		63		0.049				34.11
ATOM	424		ASP		63		0.300		4.002		44.13
ATOM	425	N	ALA		64		3.089				15.36
ATOM	426	CA	ALA		64		4.054	-5.725			17.14
ATOM	427	С	ALA		64		4.118	-5.780			21.10
ATOM	428	0	ALA		64		5.193				23.12
ATOM	429	СВ	ALA		64		5.458				20.45
ATOM	430	N	ALA		65		2.946	-6.009	4.006		22.21
ATOM	431	CA	ALA	A	65	1:	2.817	-6.072	2.565	1.00	21.81

MOTA	432	С	ALA	Α	65	13.143	-4.857	1.745	1.00 21.76
ATOM	433	0	ALA	A	65	12.855	-3.801	2.229	1.00 23.60
ATOM	434	CB	ALA	Α	65	11.384	-6.390	2.364	1.00 17.31
ATOM	435	N	LEU		66	13.401	-4.866	0.402	1.00 21.48
ATOM	436	CA	LEU		66	13.763	-3.581	-0.216	1.00 13.20
	437	C	LEU		66	12.469		-0.452	1.00 13.20
ATOM							-2.913		
MOTA	438	0	LEU		66	12.548	-1.767	-0.197	1.00 11.85
ATOM	439	CB	LEU		66	14.593	-3.602	-1.470	1.00 3.92
ATOM	440	CG	LEU		66	15.891	-4.308	-1.191	1.00 9.05
ATOM	441	CD1	LEU		66	16.509	-4.725	-2.438	1.00 12.78
ATOM	442	CD2	LEU	Α	66	16.569	-3.119	-0.580	1.00 13.44
ATOM	443	N	ALA	Α	67	11.413	-3.625	-0.801	1.00 14.94
ATOM	444	CA	ALA	A	67	10.253	-2.759	-1.277	1.00 12.42
ATOM	445	С	ALA	A	67	9.626	-1.879	-0.224	1.00 14.21
ATOM	446	0	ALA		67	9.218	-0.818	-0.643	1.00 14.29
ATOM	447	CB	ALA		67	9.089	-3.588	-1.781	1.00 3.90
ATOM	448	N	THR		68	9.494	-2.409	1.006	1.00 12.11
ATOM	449	CA	THR		68	8.780	-1.647	1.997	1.00 11.77
ATOM	450	C	THR		68	9.242	-0.214	2.219	1.00 11.77
ATOM	451	0	THR		68	8.597	0.683	2.766	1.00 11.13
ATOM	452	CB	THR		68	8.892	-2.488	3.241	1.00 13.93
ATOM	453	OG1	THR		68	10.145	-3.150	3.224	1.00 27.44
ATOM	454	CG2	THR		68	7.783	-3.459	3.087	1.00 13.39
ATOM	455	N	ASN		69	10.450	-0.057	1.808	1.00 7.59
ATOM	456	CA	ASN	Α	69	11.020	1.236	1.791	1.00 8.76
ATOM	457	С	ASN	Α	69	10.095	2.165	1.047	1.00 10.28
ATOM	458	0	ASN	Α	69	9.950	3.345	1.305	1.00 5.30
ATOM	459	CB	ASN	Α	69	12.461	1.251	1.231	1.00 5.54
ATOM	460	CG	ASN	Α	69	13.374	1.207	2.398	1.00 15.08
ATOM	461		ASN		69	13.307	2.124	3.275	1.00 31.90
ATOM	462		ASN		69	14.048	0.099	2.360	1.00 4.51
ATOM	463	N		A	70	9.390	1.656	0.079	1.00 19.09
ATOM	464	CA		A	70		2.619	-0.631	1.00 21.80
ATOM						8.552			
	465	С		A	70	7.157	2.836	-0.123	1.00 23.36
ATOM	466	0		A	70	6.509	3.717	-0.724	1.00 25.74
ATOM	467	CB		A	70	8.547	2.386	-2.082	1.00 17.38
ATOM	468	CG		Α	70	9.870	2.360	-2.770	1.00 15.72
ATOM	469		PHE	A	70	10.080	3.430	-3.576	1.00 5.15
ATOM	470	CD2	PHE	Α	70	10.702	1.245	-2.497	1.00 7.61
ATOM	471	CE1	PHE	A	70	11.268	3.330	-4.191	1.00 16.05
ATOM	472	CE2	PHE	Α	70	11.913	1.267	-3.168	1.00 22.23
ATOM	473	CZ	PHE	Α	70	12.199	2.314	-4.016	1.00 9.57
MOTA	474	N	LEU	Α	71	6.765	2.246	1.034	1.00 25.53
ATOM	475	CA	LEU		71	5.506	2.725	1.599	1.00 24.24
ATOM	476	C	LEU		71	5.649	4.037	2.343	1.00 27.91
ATOM	477	Ö	LEU		71	6.694	4.521	2.750	1.00 28.86
ATOM	478	ÇВ	LEU		71	5.150	1.635	2.730	1.00 20.00
ATOM	479	CG	LEU		71				
AT ON	413	CG	٧عب	А	<i>,</i> T	5.003	0.342	1.873	1.00 16.09

MOTA	480		LEU		71	4.879	-0.764	2.885	1.00	18.12
ATOM	481	CD2	LEU	Α	71	3.786	0.546	1.000	1.00	18.24
ATOM	482	N	PRO	Α	72	4.535	4.663	2.529	1.00	
ATOM	483	CA	PRO	A	72	4.389	5.888	3.311		34.96
MOTA	484	С	PRO	Α	72	4.865	5.590	4.778		32.90
MOTA	485	0	PRO	A	72	4.619	4.512	5.331		28.55
MOTA	486	CB	PRO		72	2.983	6.453	3.095		32.98
ATOM	487	CG	PRO		72	2.224	5.189	2.827		30.36
ATOM	488	CD	PRO		72	3.188	4.093	2.380		33.56
ATOM	489	N	ARG		73	5.601	6.610	5.221		27.54
ATOM	490	CA	ARG		73	6.325	6.547	6.408		25.42
ATOM	491	С	ARG		73	7.613	5.755	6.321		21.78
ATOM	492	ō	ARG		73	8.360	5.950	7.304		29.61
ATOM	493	СВ	ARG		73	5.469	5.978	7.549		24.29
ATOM	494	CG	ARG		73	4.575	6.998	8.155		23.47
ATOM	495	CD	ARG		73	3.818	6.793	9.360		29.73
ATOM	496	NE	ARG		73	3.222	5.460	9.392		
ATOM	497	CZ	ARG		73	2.891	5.312			36.30
ATOM	498	NH1			73	3.145	6.288	10.713		42.26
ATOM	499	NH2			73	2.320		11.555		26.57
ATOM	500	N	GLY		74		4.144 4.909	10.883		39.03
ATOM	501	CA	GLY		74	7.868 9.120		5.326	1.00	8.42
ATOM	502	C	GLY		74		4.291	5.332	1.00	5.06
ATOM	503	0	GLY		74	9.243	2.858	5.508		12.74
ATOM	504	N	THR		75	10.256	2.286	5.317		16.46
ATOM	505	CA	THR		75	8.145	2.321	5.906		12.82
ATOM	506	CA	THR		75	8.036	0.869	6.008		11.14
ATOM	507	0	THR		75	6.625	0.428	6.134		10.64
ATOM	508	CB	THR			5.757	1.231	5.949	1.00	9.36
ATOM	509	OG1	THR		75 75	8.843	0.398	7.219	1.00	6.97
ATOM	510	CG2				8.938	-0.950	7.125	1.00	5.64
ATOM	511		THR		75	8.108	0.865	8.603	1.00	6.30
		N	SER		76 76	6.409	-0.858	6.259		10.07
ATOM	512	CA	SER		76	5.061	-1.384	6.354		13.33
ATOM	513	C	SER		76	4.405	-1.163	7.747		21.87
ATOM	514	0	SER		76	5.228	-1.102	8.679		24.22
ATOM	515	CB	SER		76	5.030	-2.832	6.083	1.00	4.81
ATOM	516	OG	SER		76	5.327	-3.664	7.107		16.98
ATOM	517	N	GLN		77	3.082	-1.100	7.911		24.90
ATOM	518	CA	GLN		77	2.454	-1.020	9.166		23.85
ATOM	519	С	GLN		77	2.643	-2.236	10.015		19.58
ATOM	520	0	GLN		77	2.908	-2.140	11.203		15.15
MOTA	521	CB	GLN		77	0.983	-0.703	9.217	1.00	32.64
ATOM	522	CG	GLN		77	0.567	-0.580	10.642	1.00	49.56
ATOM	523	CD	GLN		77	0.689	0.785	11.194	1.00	65.91
MOTA	524	OE1			77	0.956	0.869	12.356	1.00	66.06
ATOM	525		GLN		77	0.481	1.750	10.350	1.00	68.91
ATOM	526	N	ALA		78	2.754	-3.376	9.402	1.00	15.90
ATOM	527	CA	ALA	A	78	3.071	-4.577	10.073	1.00	19.47

ATOM	528	С	ALA		78	4.381	-4.332	10.819	1.00 24.48
ATOM	529	0	ALA		78	4.389	-4.729	11.983	1.00 26.91
ATOM	530	CB	ALA	Α	78	3.390	-5.808	9.336	1.00 17.23
ATOM	531	N	ASN	Α	79	5.350	-3.863	10.093	1.00 21.58
ATOM	532	CA	ASN	Α	79	6.602	-3.576	10.774	1.00 20.62
ATOM	533	С	ASN	Α	79	6.480	-2.673	11.969	1.00 20.93
ATOM	534	0	ASN	Α	79	6.975	-2.944	13.053	1.00 15.52
ATOM	535	CB	ASN		79	7.474	-3.069	9.670	1.00 24.79
ATOM	536	CG	ASN		79	7.933	-4.238	8.824	1.00 28.76
ATOM	537		ASN		79	7.867	-5.439	9.091	1.00 25.30
ATOM	538	ND2			79	8.488	-3.891	7.660	1.00 24.90
ATOM	539	N	ILE		80	5.731	-1.611	11.936	1.00 24.90
ATOM	540	CA	ILE		80	5.586	-0.574	12.924	
ATOM	541	C	ILE		80				1.00 17.00
						4.925	-1.187	14.118	1.00 20.63
ATOM	542	0	ILE		80	5.234	-0.939	15.264	1.00 18.79
ATOM	543	CB	ILE		80	4.756	0.629	12.436	1.00 11.98
ATOM	544	CG1	ILE		80	5.627	1.124	11.297	1.00 9.50
ATOM	545	CG2	ILE		80	4.379	1.728	13.354	1.00 16.27
ATOM	546	CD1	ILE		80	5.007	2.071	10.424	1.00 8.15
ATOM	547	N	ASP		81	4.017	-2.019	13.708	1.00 19.21
ATOM	548	CA	ASP		81	3.304	-2.778	14.728	1.00 15.15
ATOM	549	С	ASP	A	81	4.147	-3.711	15.510	1.00 15.77
ATOM	550	0	ASP	Α	81	4.084	-3.697	16.695	1.00 15.82
ATOM	551	CB	ASP	Α	81	2.291	-3.438	13.868	1.00 26.36
ATOM	552	CG	ASP	Α	81	1.065	-2.530	13.790	1.00 23.71
ATOM	553	OD1	ASP	Α	81	1.105	-1.355	14.226	1.00 14.33
ATOM	554	OD2	ASP	Α	81	0.061	-3.125	13.222	1.00 33.05
ATOM	555	N	GLU	Α	82	5.148	-4.447	15.096	1.00 16.07
ATOM	556	CA	GLU	Α	82	5.984	-5.318	15.882	1.00 14.77
ATOM	557	С	GLU		82	6.839	-4.355	16.667	1.00 19.33
ATOM	558	0	GLU		82	7.315	-4.708	17.752	1.00 23.58
ATOM	559	СВ	GLU		82	6.998	-6.031	15.064	1.00 13.20
ATOM	560	CG	GLU		82	7.792	-7.239	15.476	1.00 23.09
ATOM	561	CD	GLU		82	6.767	-8.114	16.185	1.00 29.68
ATOM	562	OE1	GLU		82	5.666	-7.670	16.403	1.00 25.68
ATOM	563	OE1	GLU		82				
ATOM	564	N			83	7.273	-9.181	16.411	1.00 33.08
			GLY			7.228	-3.227	16.199	1.00 16.79
ATOM	565	CA	GLY		83	8.033	-2.428	17.140	1.00 17.32
ATOM	566	C	GLY		83	7.238	-2.018	18.366	1.00 17.54
ATOM	567	0	GLY		83	7.561	-2.103	19.528	1.00 15.06
ATOM	568	N	LYS		84	6.093	-1.408	18.114	1.00 18.72
ATOM	569	CA		A	84	5.050	-1.146	19.096	1.00 16.90
ATOM	570	С		Α	84	4.893	-2.337	20.057	1.00 17.74
ATOM	571	0	LYS	A	84	4.962	-2.265	21.295	1.00 14.31
MOTA	572	CB	LYS	Α	84	3.799	-0.872	18.307	1.00 14.62
ATOM	573	CG	LYS	Α	84	3.535	0.565	18.291	1.00 19.30
ATOM	574	CD	LYS	A	84	2.787	1.013	17.044	1.00 34.24
ATOM	575	CE	LYS	Α	84	1.568	1.902	17.337	1.00 37.70

ATOM	576	NZ	LYS	7\	84	0.346	1.226	16.827	1 00	48.42
ATOM	577	N	ARG		85	4.617	-3.506	19.519	1.00	
	•	CA						20.280		
ATOM	578		ARG		85	4.583	-4.705		1.00	
ATOM	579	C	ARG		85	5.677	-4.733	21.308	1.00	
ATOM	580	0 .	ARG		85	5.442	-5.192	22.383	1.00	
ATOM	581	CB	ARG		85	4.740	-5.979	19.464	1.00	
ATOM	582	CG	ARG		85	3.843	-7.094	19.887	1.00	8.85
ATOM	583	CD	ARG		85	4.146	-8.554	19.705	1.00	7.20
ATOM	584	NE	ARG		85	5.483	-8.898	19.194	1.00	
ATOM	585	CZ	ARG	Α	85	6.170	-9.705	19.899	1.00	
ATOM	586	NH1	ARG	Α	85	5.627	-10.161	21.040	1.00	34.03
ATOM	587	NH2	ARG	Α	85	7.345	-9.979	19.555	1.00	15.36
ATOM	588	N	LEU	A	86	6.901	-4.586	20.956	1.00	22.21
ATOM	589	CA	LEU	Α	86	8.006	-4.792	21.873	1.00	20.94
ATOM	590	C.	LEU	Α	86	8.044	-3.637	22.803	1.00	20.73
ATOM	591	0	LEU		86	8.155	-3.970	23.925		22.18
ATOM	592	CB	LEU		86	9.333	-4.932	21.168	1.00	6.67
ATOM	593	CG	LEU		86	9.358	-6.241	20.282	1.00	
ATOM	594	CD1			86	10.546	-6.054	19.287	1.00	
ATOM	595	CD2	LEU		86	9.362	-7.516	21.020	1.00	5.17
ATOM	596	N		A	87	7.700	-2.446	22.529	1.00	
ATOM	597	CA		A	87	7.700	-2.446	23.492	1.00	18.21
	598	C					-1.410		1.00	
MOTA				A	87	6.939		24.618 25.839		
ATOM	599	0		A	87	7.082	-1.565		1.00	30.36
ATOM	600	CB		A	87	7.498	-0.118	22.846		15.81
ATOM	601	CG	PHE	A	87	8.661	0.503	22.128		22.72
ATOM	602	CD1		A	87	9.625	1.163	22.795		25.90
MOTA	603	CD2		A	87	8.800	0.446	20.774		24.19
ATOM	604	CE1		A	87	10.699	1.781	22.220		26.46
ATOM	605	CE2		A	87	9.871	0.991	20.153		29.24
ATOM	606	CZ	PHE	Α	87	10.827	1.669	20.849	1.00	20.81
MOTA	607	N	ALA	A	88	5.862	-2.422	24.266	1.00	29.15
ATOM	608	CA	ALA	Α	88	4.772	-2.699	25.195	1.00	22.92
ATOM	609	С	ALA	Α	88	5.186	-3.837	26.068	1.00	22.03
ATOM	610	0	ALA	Α	88	4.974	-3.879	27.284	1.00	27.02
ATOM	611	CB	ALA	Α	88	3.551	-2.803	24.299	1.00	22.13
ATOM	612	N	LEU		89	5.649	-4.897	25.531		19.16
ATOM	613	CA	LEU		89	6.188	-6.032	26.208		19.29
ATOM	614	C	LEU		89	7.250	-5.507	27.133		22.06
ATOM	615	0	LEU		89	7.449	-6.050	28.177		20.49
ATOM	616	CB	LEU		89	7.021	-6.863	25.221		18.41
ATOM	617	CG	LEU		89	7.477	-8.167	25.834		20.45
ATOM	618		LEU		89	6.326	-8.707	26.627		17.22
	619		LEU		89					18.83
ATOM						8.060	-9.057	24.769		
ATOM	620	N	ALA		90	8.124	-4.644	26.722		22.80
ATOM	621	CA	ALA		90	9.027	-4.137	27.701		24.14
ATOM	622	C	ALA		90	8.237	-3.488	28.849		23.63
ATOM	623	0	ALA	A	90	8.414	-3.835	30.071	1.00	22.73

ATOM	624	CB	ALA		90	10.080	-3.253	27.139	1.00	7.74
ATOM	625	N	ASN	Α	91	7.457	-2.445	28.732	1.00	25.45
MOTA	626	CA	ASN	Α	91	6.665	-1.979	29.870	1.00	27.25
ATOM	627	С	ASN	Α	91	5.847	-2.996	30.656	1.00	30.97
ATOM	628	0	ASN	Α	91	5.346	-2.884	31.768	1.00	27.64
ATOM	629	CB	ASN		91	5.560	-1.206	29.125	1.00	29.14
ATOM	630	CG	ASN		91	4.946	-0.345	30.216	1.00	31.73
ATOM	631	OD1			91	3.845	-0.692	30.645		46.76
ATOM	632		ASN		91	5.641	0.629	30.643	1.00	
ATOM	633	N	GLN		92	5.369	-4.008	29.969		35.37
ATOM	634	CA	GLN		92	4.702	-5.141	30.591		35.55
	635	C	GLN		92	5.619	-6.072	31.352		34.28
ATOM										
ATOM	636	0	GLN		92	5.227	-6.519	32.440	1.00	39.47
ATOM	637	CB	GLN		92	3.866	-5.903	29.573		54.94
MOTA	638	CG	GLN		92	2.689	-6.698	30.142		78.63
MOTA	639	CD	GLN		92	2.806	-8.167	29.805	1.00	
MOTA	640	OE1			92	3.597	-8.840	30.475		96.99
ATOM	641	NE2	GLN		92	2.083	-8.696	28.824	1.00	97.81
ATOM	642	N	LYS	Α	93	6.859	-6.403	31.050	1.00	31.97
ATOM	643	CA	LYS	A	93	7.675	-7.204	31.972	1.00	25.22
ATOM	644	С	LYS	Α	93	8.381	-6.298	33.015	1.00	24.68
ATOM	645	0	LYS	Α	93	8.716	-6.793	34.075	1.00	32.13
ATOM	646	CB	LYS	Α	93	8.673	-7.980	31.148	1.00	10.86
ATOM	647	CG	LYS	A	93	8.225	-8.963	30.159	1.00	24.26
ATOM	648	CD	LYS	Α	93	9.362	-9.966	29.986	1.00	21.96
ATOM	649	CE	LYS		93		-10.718	28.658		23.78
ATOM	650	NZ	LYS		93		-11.805	28.300		25.87
ATOM	651	N	CYS		94	8.752	-5.096	32.774	1.00	16.62
ATOM	652	CA	CYS		94	9.752	-4.412	33.480	1.00	18.95
ATOM	653	C	CYS		94	9.512	-2.936	33.537		24.83
ATOM	654	0	CYS		94	10.184	-2.017	33.150	1.00	26.80
ATOM	655	CB		A	94	11.147	-4.691	32.911	1.00	3.14
ATOM		SG			94					
	656		CYS			11.618	-6.437	32.882		25.28
ATOM	657	N	PRO		95	8.403	-2.561	34.086		26.08
ATOM	658	CA	PRO		95	7.891	-1.202	33.878	1.00	26.11
ATOM	659	С	PRO		95	8.960	-0.259	34.299	1.00	27.32
MOTA	660	0	PRO		95	8.776	0.966	34.108	1.00	29.08
MOTA	661	CB	PRO		95	6.609	-1.090	34.747		20.75
ATOM	662	CG	PRO		95	6.587	-2.421	35.322		19.04
ATOM	663	CD	PRO	A	95	7.363	-3.461	34.509	1.00	22.55
ATOM	664	N	ASN	Α	96	9.836	-0.776	35.193	1.00	31.44
MOTA	665	CA	ASN		96	10.559	0.274	35.966	1.00	35.38
MOTA	666	С	ASN	Α	96	11.891	0.476	35.353	1.00	33.83
MOTA	667	0	ASN		96	12.599	1.359	35.684	1.00	33.31
MOTA	668	CB	ASN		96	10.558	-0.099	37.429	1.00	53.70
ATOM	669	CG	ASN		96	9.238	0.342	38.026	1.00	
ATOM	670		ASN		96	8.758	1.432	37.706	1.00	
ATOM	671		ASN		96	8.676	-0.526	38.861	1.00	
					- 0	3.070				520

ATOM	672	NT	מנות	70	07	10 007	0 400	24 505		
ATOM	673	N	THR		97	12.287	-0.409	34.507		30.32
		CA	THR		97	13.519	-0.367	33.794		22.83
ATOM	674	С	THR		97	13.404	0.493	32.534		22.44
ATOM	675	0	THR		97	12.446	0.779	31.816		21.14
ATOM	676	CB	THR		97	13.835	-1.851	33.705		25.87
ATOM	677	OG1			97	14.602	-1.915	32.528		38.91
ATOM	678	CG2			97	12.769	-2.901	33.621		24.22
ATOM	679	N	PRO		98	14.393	1.415	32.408		20.59
ATOM	680	CA	PRO		98	14.513	2.292	31.254	1.00	18.15
ATOM	681	С	PRO		98	14.882	1.494	29.978	1.00	16.07
ATOM	682	0	PRO		98	15.622	0.462	29.934	1.00	17.19
ATOM	683	CB	PRO		98	15.563	3.339	31.676	1.00	14.55
ATOM	684	CG	PRO	Α	98	16.270	2.646	32.699	1.00	12.29
ATOM	685	CD	PRO	Α	98	15.735	1.331	33.046	1.00	12.02
ATOM	686	N	VAL	Α	99	14.322	2.107	28.940	1.00	13.81
ATOM	687	CA	VAL	Α	99	14.225	1.544	27.632		14.02
ATOM	688	С	VAL	Α	99	14.956	2.407	26.663		10.66
ATOM	689	0	VAL	Α	99	14.716	3.679	26.712	1.00	6.90
ATOM	690	CB	VAL	A	99	12.673	1.343	27.335	1.00	2.87
ATOM	691	CG1	VAL	Α	99	12.666	1.272	25.872		17.40
ATOM	692	CG2	VAL	Α	99	12.442	-0.111	27.744	1.00	5.75
ATOM	693	N	VAL	Α	100	15.885	1.776	25.861	1.00	6.45
ATOM	694	CA	VAL			16.525	2.755	24.900	1.00	9.61
ATOM	695	С	VAL			16.389	2.159	23.561		10.79
ATOM	696	0	VAL			16.256	0.973	23.477	1.00	9.11
ATOM	697	CB	VAL			17.877	3.260	25.197	1.00	8.05
ATOM	698	CG1	VAL	Α	100	17.824	4.252	26.336	1.00	6.05
ATOM	699		VAL			18.853	2.053	25.591	1.00	6.68
ATOM	700	N	ALA			16.277	2.928	22.511		13.14
ATOM	701	CA	ALA.			16.127	2.266	21.183	1.00	15.67
ATOM	702	С	ALA .			17.065	2.747	20.053	1.00	12.08
ATOM	703	0	ALA .			17.261	4.042	19.907	1.00	11.16
ATOM	704	CB	ALA			14.685	2.609	20.812	1.00	6.57
ATOM	705	N	GLY .			17.218	1.787	19.099	1.00	7.53
ATOM	706	CA	GLY .			17.949	2.415	17.939	1.00	7.10
ATOM	707	C	GLY .			17.477	1.803	16.744	1.00	7.10
ATOM	708	Ō	GLY .			17.102	0.621	16.878		10.83
ATOM	709	N	GLY :			17.706	2.407	15.648		
ATOM	710	CA	GLY .			17.446	1.745		1.00	7.80
ATOM	711	C	GLY :			18.303	2.211	14.356	1.00	5.33
ATOM	712	0	GLY :			18.785		13.180	1.00	7.56
ATOM	713	N	TYR .				3.340	13.227	1.00	6.88
ATOM	714	CA	TYR :			18.490	1.387	12.139	1.00	7.09
ATOM	715	C	TYR			19.392	1.682	11.069	1.00	5.99
ATOM	716	0	TYR :			18.705	1.614	9.705	1.00	9.47
ATOM	717	CB				18.115	0.638	9.441	1.00	6.46
ATOM	718	CG	TYR			20.592	0.797	11.079	1.00	5.40
ATOM	719		TYR			21.436	1.078	9.876	1.00	8.05
FITOR	113	CD1	TYR I	~	104	21.708	2.302	9.352	1.00	5.91

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ATOM	720		TYR			21.961	-0.044	9.172	1.00	6.85
ATOM	721	CE1	TYR			22.447	2.513	8.186	1.00	5.61
ATOM	722	CE2	TYR	A	104	22.751	0.052	8.072	1.00	7.49
ATOM	723	CZ	TYR	Α	104	22.972	1.377	7.608	1.00	11.08
ATOM	724	OH	TYR	Α	104	23.795	1.509	6.479	1.00	14.32
ATOM	725	N	SER	A	105	18.939	2.975	8.852	1.00	18.39
ATOM	726	CA	SER			18.190	2.854	7.601	1.00	9.66
ATOM	727	С	SER			16.763	2.370	7.722	1.00	6.10
ATOM	728	Ō	SER			16.090	3.304	8.077	1.00	5.63
ATOM	729	CB	SER			19.124	2.159	6.607	1.00	8.55
ATOM	730	OG	SER			18.553	1.685	5.463		24.30
	731	N								
ATOM			GLN			16.241	1.405	7.079	1.00	9.93
ATOM	732	CA	GLN			14.759	1.316	7.002	1.00	8.25
ATOM	733	С	GLN			14.453	1.089	8.473	1.00	8.51
ATOM	734	0	GLN			13.470	1.683	8.862	1.00	6.31
ATOM	735	CB	GLN			14.239	0.393	5.940	1.00	7.45
ATOM	736	CG	GLN			13.184	-0.528	6.465	1.00	18.04
ATOM	737	CD	GLN	Α	106	12.228	-1.220	5.581	1.00	16.87
ATOM	738	OE1	GLN	Α	106	11.024	-1.180	5.492	1.00	17.59
ATOM	739	NE2	GLN .	A	106	12.643	-2.032	4.713	1.00	8.32
ATOM	740	N	GLY .	Α	107	15.269	0.310	9.172	1.00	7.13
ATOM	741	CA	GLY .			15.190	0.159	10.606	1.00	4.61
ATOM	742	С	GLY .			15.048	1.472	11.356	1.00	8.27
ATOM	743	Ō	GLY			14.219	1.511	12.290	1.00	6.52
ATOM	744	N	ALA.			15.653	2.637	11.033	1.00	6.44
ATOM	745	CA	ALA .			15.266	3.864	11.641	1.00	7.41
ATOM	746	C	ALA			13.813	4.346	11.471		11.76
ATOM	747	0	ALA .			13.150	4.914	12.298		12.64
ATOM	748	CB	ALA			16.121	5.006	11.170		13.93
ATOM	749	N	ALA .			13.321	4.312	10.267	1.00	9.78
ATOM	750	CA	ALA .			12.056	4.685	9.861		10.47
ATOM	751	С	ALA .			11.093	3.858	10.727		12.32
ATOM	752	0	ALA .			10.016	4.391	11.035		14.67
ATOM	753	CB	ALA.			12.035	4.173	8.456	1.00	10.24
MOTA	754	N	LEU .	A	110	11.259	2.690	11.077	1.00	4.34
ATOM	755	CA	LEU .	Α	110	10.458	1.760	11.783	1.00	11.71
ATOM	756	С	LEU .	A	110	10.305	2.253	13.203	1.00	15.26
ATOM	757	0	LEU .	Α	110	9.298	2.672	13.685	1.00	18.07
ATOM	758	CB	LEU .				0.319	11.634	1.00	
ATOM	759	CG	LEU			10.247	-0.801	12.258	1.00	
ATOM	760		LEU .			10.685	-2.233	11.862	1.00	
ATOM	761	CD2				10.278	-0.659	13.783	1.00	
ATOM	762	N	ILE .			11.397		13.703		15.77
ATOM	763	CA	ILE .				2.373	15.246		12.22
ATOM	764	C	ILE .			11.027		15.234	1.00	
ATOM	765	O	ILE .			10.404		16.241		12.54
ATOM	766	CB	ILE .			12.977				15.55
ATOM	767	CGI	ILE .	A	TII	13.222	1.279	15.805	1.00	14.19

ATOM	768	CG2	ILE	Α	111	13.	. 195	3.	465	17.0	05	1.00	4.64
MOTA	769	CD1	ILE	Α	111	12.	.410	0.	887	17.0	02	1.00	14.88
ATOM	770	N	ALA	Α	112	11.	. 309	5.	170	14.3	41	1.00	11.00
ATOM	771	CA	ALA	Α	112	10.	. 792	6.	528	14.4	27	1.00	12.45
ATOM	772	С	ALA	A	112		.266		455	14.3		1.00	
ATOM	773	0			112		728		131	15.1		1.00	
ATOM	774	СВ			112		334		505	13.4		1.00	5.70
ATOM	775	N			113		575		572	13.5		1.00	
ATOM	776	CA			113		167		512	13.5		1.00	15.39
ATOM	777	C			113		475		093	14.8		1.00	18.21
ATOM	778	0			113		498		750	15.2		1.00	
ATOM	779	CB			113					12.5			14.59
							678		562			1.00	17.63
ATOM	780	N			114		937		948	15.3		1.00	16.02
ATOM	781	CA.	ALA				483		218	16.4		1.00	16.43
ATOM	782	C	ALA				578		114	17.6			22.20
ATOM	783	0	ALA				673		321	18.4		1.00	18.94
ATOM	784	CB	ALA				474		084	16.5		1.00	4.69
ATOM	785	N	VAL				722		836	17.7			22.46
ATOM	786	CA	VAL			7.	855	5.	499	19.0		1.00	20.88
ATOM	787	С	VAL			6.	670	6.	469	19.0		1.00	22.71
ATOM	788	Ο.	VAL			6.	136	6.	761	20.0	57	1.00	22.05
ATOM	789	CB	VAL	Α	115	9.	279	6.	090	19.1	37	1.00	19.61
ATOM	790	CG1	VAL	Α	115	9.	396	7.	259	20.1	22	1.00	8.35
ATOM	791	CG2	VAL	Α	115	10.	245	5.	016	19.5	62	1.00	13.91
ATOM	792	N	SER	Α	116	6.	467	7.	085	17.83	28	1.00	23.59
ATOM	793	CA	SER	A	116	5.	539	8.	172	17.7	36	1.00	23.68
ATOM	794	С	SER	A	116	4.	169	7.	647	18.1	20	1.00	23.77
ATOM	795	0	SER	A	116	3.	333	8.	523	18.3	99		27.35
ATOM	796	CB	SER	Α	116		522		865	16.3			25.21
ATOM	797	OG	SER				168		043	15.2			28.05
ATOM	798	N	GLU				859		397	18.0			18.83
ATOM	799	CA	GLU				491		020	18.2			22.21
ATOM	800	C	GLU				461		474	19.6			30.46
ATOM	801	ō	GLU				487		773	19.8			35.72
ATOM	802	СВ	GLU				977		902	17.3		1.00	21.63
ATOM	803	CG	GLU				167		219	15.89			26.41
ATOM	804	CD	GLU				560		424	14.83			
ATOM	805		GLU				912		440				34.01
ATOM	806		GLU							15.04			32.59
ATOM	807						750		833	13.6			44.62
		N C7	LEU				438		570	20.5			34.45
ATOM	808	CA	LEU				326		006	21.83			33.64
ATOM	809	С	LEU				681		110	22.63			41.75
ATOM	810	0	LEU				594		267	22.3			39.90
ATOM	811	CB	LEU				600		668	22.39			29.44
ATOM	812	CG	LEU				628		391	21.6			26.36
ATOM	813		LEU				921		340	22.3			27.53
ATOM	814		LEU				110		520	21.5			20.69
ATOM	815	N	SER	Α	119	2.	076	5.	794	23.72	26	1.00	48.86

7 TOM	816	CA	SER	7	110	0.910	5.647	24.476	1 00	52.44
ATOM								- · ·		
ATOM	817	C	SER			1.212	6.063	25.866		52.57
ATOM	818	0	SER			1.485	5.258	26.735	1.00	
ATOM	819	CB	SER			0.550	4.132	24.488	1.00	
MOTA	820	OG	SER			1.393	3.091	23.908	1.00	
MOTA	821	N	GLY	Α	120	1.532	7.307	26.024		52.95
ATOM	822	CA	GLY	A	120	1.910	7.761	27.382		53.35
ATOM	823	С	GLY	Α	120	2.944	7.109	28.291	1.00	49.09
ATOM	824	0	GLY	A	120	4.086	7.617	28.358	1.00	49.66
ATOM	825	N	ALA	Α	121	2.526	6.129	29.102	1.00	42.97
ATOM	826	CA	ALA			3.477	5.574	30.022	1.00	40.72
ATOM	827	C	ALA			4.587	4.772	29.326	1.00	44.20
ATOM	828	Ō	ALA			5.749	4.803	29.711		45.42
ATOM	829	СВ	ALA			2.965	4.542	30.903		36.34
ATOM	830	N	VAL			4.122	4.035	28.312		41.15
	831	CA	VAL			5.090	3.269	27.548	1.00	
ATOM								26.652		28.48
ATOM	832	C	VAL			5.870	4.168			
ATOM	833	0	VAL			7.084	4.019	26.872		27.69
ATOM	834	CB	VAL			4.424	2.056	26.952		30.22
ATOM	835	CG1	VAL			2.924	1.997	27.098		28.03
ATOM	836	CG2	VAL			4.891	1.836	25.551		23.22
ATOM	837	N	LYS			5.424	5.310	26.177		23.16
ATOM	838	CA	LYS			6.354	6.314	25.661		23.11
MOTA	839	С	LYS			7.403	6.783	26.661		25.28
ATOM	840	0	LYS	Α	123	8.524	7.224	26.449		29.01
ATOM	841	CB	LYS	Α	123	5.561	7.502	25.100		23.54
ATOM	842	CG	LYS	Α	123	6.171	8.573	24.277	1.00	26.71
ATOM	843	CD	LYS	Α	123	5.400	9.775	23.888	1.00	43.07
ATOM	844	CE	LYS	Α	123	4.953	9.783	22.461	1.00	59.59
ATOM	845	NZ	LYS	Α	123	3.518	9.637	22.099	1.00	67.50
ATOM	846	N	GLU	A	124	6.977	6.991	27.918	1.00	27.95
ATOM	847	CA	GLU			7.845	7.700	28.863	1.00	27.29
ATOM	848	С			124	8.910	6.706	29.243		25.21
ATOM	849	Ō	GLU			9.993	7.165	29.769	1.00	
ATOM	850	СВ	GLU			6.986	8.351	29.927		40.13
ATOM	851	CG			124	7.588	8.609	31.295	1.00	
ATOM	852	CD	GLU			8.530	9.814	31.247		66.99
								31.902		70.44
ATOM	853		GLU			9.619	9.751			73.84
ATOM	854		GLU			7.949	10.652	30.502		
ATOM	855	N	GLN			8.656	5.393	29.058		19.93
MOTA	856	CA			125	9.761	4.509	29.546		17.98
ATOM	857	С	GLN			10.865	4.556	28.521		24.28
ATOM	858	0			125	11.964	4.107	28.815		21.47
ATOM	859	CB			125	9.225	3.178	29.844	1.00	9.13
MOTA	860	CG	GLN			9.901	2.001	30.299	1.00	9.05
MOTA	861	CD			125	9.211	0.719	30.129	1.00	
MOTA	862	OE1	GLN	A	125	8.190	0.703	29.466		28.52
MOTA	863	NE2	GLN	Α	125	9.662	-0.396	30.684	1.00	13.34

ATOM	864	N			126	10.593	5.188	27.319	1.00	25.30
ATOM	865	CA	VAL	A	126	11.738	5.124	26.361	1.00	22.55
ATOM	866	С	VAL	Α	126	12.546	6.334	26.614	1.00	17.55
ATOM	867	0	VAL	A	126	12.109	7.408	26.329	1.00	12.79
ATOM	868	CB	VAL	Α	126	11.227	4.560	25.022	1.00	23.76
ATOM	869	CG1	VAL	Α	126	9.706	4.686	24.946	1.00	23.77
ATOM	870	CG2	VAL	A	126	11.795	5.081	23.743	1.00	23.81
ATOM	871	N	LYS	Α	127	13.726	6.233	27.264	1.00	
ATOM	872	CA	LYS	Α	127	14.462	7.494	27.639	1.00	18.18
ATOM	873	С			127	15.239	8.063	26.488	1.00	
ATOM	874	0			127	15.812	9.103	26.680	1.00	
ATOM	875	СВ			127	15.401	7.148	28.792	1.00	
ATOM	876	CG			127	14.770	6.110	29.713	1.00	
MOTA	877	CD			127	13.435	6.726	30.064		33.86
ATOM	878	CE			127	12.779	6.612	31.399	1.00	
ATOM	879	NZ			127	12.279	7.863	31.993	1.00	
ATOM	880	N			128	15.522	7.281	25.416	1.00	
ATOM	881	CA			128	16.280	7.231	24.306	1.00	
ATOM	882	C			128	16.358	7.104			
ATOM	883	0			128			23.063	1.00	
ATOM	884					16.168	5.901	23.226	1.00	
	885	N			129 129	16.451	7.725	21.892	1.00	
ATOM		CA				16.497	6.872	20.691		13.82
ATOM	886	C			129	17.519	7.371	19.719	1.00	8.35
ATOM	887	0			129	17.602	8.553	19.556	1.00	3.85
ATOM	888	CB	VAL			15.192	6.426	20.054	1.00	
ATOM	889	CG1	VAL			14.007	7.041	20.726	1.00	6.50
ATOM	890	CG2	VAL			15.051	6.729	18.571	1.00	10.03
ATOM	891	N	ALA			18.455	6.398	19.363	1.00	8.05
ATOM	892	CA	ALA			19.430	6.845	18.344	1.00	7.55
ATOM	893	С	ALA			19.078	6.293	16.958	1.00	11.17
ATOM	894	0	ALA			18.755	5.145	16.849	1.00	15.74
MOTA	895	CB	ALA			20.781	6.391	18.603	1.00	5.89
ATOM	896	N	LEU			18.911	6.953	15.892	1.00	7.36
ATOM	897	CA	LEU	Α	131	18.635	6.625	14.553	1.00	7.70
ATOM	898	С	LEU	Α	131	19.876	6.908	13.661	1.00	12.02
MOTA	899	0	LEU	A	131	20.436	8.033	13.604	1.00	6.80
ATOM	900	CB	LEU	Α	131	17.604	7.713	14.102	1.00	8.40
ATOM	901	CG	LEU	Α	131	16.160	7.830	14.575	1.00	6.67
ATOM	902	CD1	LEU	Α	131	15.391	8.957	13.981	1.00	4.49
ATOM	903	CD2	LEU			15.481	6.488	14.324	1.00	5.12
ATOM	904	N	PHE			20.271	6.009	12.802	1.00	
ATOM	905	CA	PHE			21.422	6.183	11.908		10.44
ATOM	906	С	PHE			20.965	6.013	10.478	1.00	8.46
ATOM	907	Ō	PHE			20.175	5.101	10.097		11.04
ATOM	908	CB	PHE			22.217	4.931	12.282		10.56
ATOM	909	CG	PHE			22.693	4.830	13.714		16.38
ATOM	910	CD1	PHE			21.951	4.029	14.542		13.36
ATOM	911		PHE			23.860	5.489	14.213		15.12
		- D- L		11		23.000	J.403	13.710	1.00	10.12

ATOM	912		PHE			22.342	3.911	15.889	1.00	14.91
ATOM	913	CE2	PHE	Α	132	24.176	5.323	15.513		18.02
ATOM	914	CZ	PHE	Α	132	23.426	4.530	16.403	1.00	
ATOM	915	N	GLY	Α	133	21.431	6.876	9.580	1.00	
ATOM	916	CA	GLY	A	133	21.026	6.893	8.148	1.00	
ATOM	917	С	GLY	Α	133	19.503	6.919		1.00	
ATOM	918	0			133	18.890	5.926	7.593	1.00	
ATOM	919	N			134	18.926	8.070	8.532	1.00	
ATOM	920	CA			134	17.455	8.022	8.838	1.00	
ATOM	921	С			134	16.647	8.365	7.584	1.00	
ATOM	922	0			134	16.785		7.131	1.00	5.85
ATOM	923	CB			134	17.161	9.128	9.836	1.00	7.27
ATOM	924	CG			134	15.842	9.393	10.391	1.00	7.89
ATOM	925	CD1			134	14.889	8.437	10.312	1.00	6.65
ATOM	926	CD2			134	15.661	10.651	10.948	1.00	
ATOM	927	CE1			134	13.657	8.690	10.821	1.00	9.05
ATOM	928	CE2			134	14.408	10.928	11.467	1.00	
ATOM	929	CZ			134	13.428	9.923	11.423	1.00	
ATOM	930	OH			134	12.146	10.110	11.975		12.41
ATOM	931	N			135	15.811	7.398	7.139	1.00	
ATOM	932	CA			135	15.229	7.581	5.789	1.00	7.71
ATOM	933	С			135	14.082	8.530	5.825	1.00	
ATOM	934	Ō			135	13.845	8.878	4.727		11.26
ATOM	935	СВ			135	14.772	6.394	4.967		12.02
ATOM	936	OG1	THR			13.821	5.399	5.398		22.81
ATOM	937	CG2	THR			15.828	5.332	4.712		14.88
MOTA	938	N	GLN			13.632	9.105	6.928		15.28
ATOM	939	CA	GLN			12.596	10.134	6.968		16.48
ATOM	940	С	GLN			13.102	11.418	7.646		17.46
MOTA	941	Ō	GLN			12.292	12.231	8.035		12.82
MOTA	942	СВ	GLN			11.336	9.671	7.701	1.00	5.71
ATOM	943	CG	GLN			11.178	8.191	7.263		13.60
ATOM	944	CD	GLN			10.504	8.264	5.932		14.65
MOTA	945	OE1				9.587	9.102	5.986		23.99
ATOM	946	NE2	GLN			10.852	7.529	4.914		14.68
ATOM	947	N	ASN			14.421	11.532	7.566		18.52
ATOM	948	CA	ASN			14.953	12.752	8.141		18.16
ATOM	949	C	ASN			14.301	13.929	7.458		19.79
ATOM	950	Ō	ASN			13.895	14.802	8.157		12.28
ATOM	951	CB	ASN			16.481	12.573	8.239		14.17
ATOM	952	CG	ASN			17.247	13.740	8.812		19.75
MOTA	953		ASN			17.821	14.341	7.934		14.52
ATOM	954		ASN			17.390	14.130	10.042		17.43
ATOM	955	N	LEU			14.180	14.130	6.141		27.31
ATOM	956	CA	LEU			13.640	15.270	5.553		25.53
ATOM	957	C	LEU			12.190				
ATOM	958	0	LEU			11.710	15.332 16.281	5.971 6.549		22.45
ATOM	959	CB	LEU			13.632				25.13
		Q <i>D</i>		4.1	100	13.032	15,269	4.056	1.00	41.28

ATOM	960	CG	LEU A	138	13.713	16.582	3.303	1.00	31.76
ATOM	961	CD1	LEU A	138	14.641	17.503	4.012		51.09
ATOM	962	CD2	LEU A	138	14.207	16.573	1.958	1.00	
MOTA	963	N	GLN A	139	11.378	14.403	5.569	1.00	
ATOM	964	CA	GLN A	139	10.034	14.390	6.037	1.00	
ATOM	965	С	GLN A		9.846	14.749	7.471	1.00	
ATOM	966	0	GLN A		8.791	15.282	7.528	1.00	
ATOM	967	CB	GLN A		9.517	12.969	5.899	1.00	
ATOM	968	CG	GLN A		9.684	12.643	4.450	1.00	
ATOM	969	CD	GLN A		10.984	11.983	4.110		22.69
ATOM	970	OE1			10.674	10.980	3.477		35.62
ATOM	971	NE2			12.195	12.405	4.410		31.70
ATOM	972	N	ASN A		10.454	14.072	8.427	1.00	
ATOM	973	CA	ASN A		10.215	14.183	9.848	1.00	
ATOM	974	С	ASN A		10.941	15.429	10.293	1.00	
ATOM	975	0	ASN A		11.040	15.654	11.454	1.00	18.05
ATOM	976	CB	ASN A		10.581	12.910	10.541	1.00	
ATOM	977	CG	ASN A		9.465	11.998	10.210	1.00	
ATOM	978	OD1			8.615	12.565	9.563		23.57
ATOM	979	ND2			9.460	10.756	10.630		22.65
ATOM	980	N	ARG A		11.457	16.162	9.397		19.20
ATOM	981	CA	ARG A		12.170	17.350	9.790		26.25
ATOM	982	С	ARG A		13.219	17.090	10.818		25.06
ATOM	983	0	ARG A		13.365	17.928	11.649		27.60
ATOM	984	CB	ARG A		11.123	18.299	10.271		37.72
ATOM	985	CG	ARG A		10.083	18.974	9.372		49.61
ATOM	986	N	GLY A		14.110	16.165	10.920		19.42
MOTA	987	CA	GLY A	142	14.997	15.778	11.902		14.21
MOTA	988	С	GLY A	142	14.652	15.066	13.158		19.42
ATOM	989	0	GLY A	142	15.547	14.759	13.971		23.74
ATOM	990	N	GLY A	143	13.354	14.851	13.569		14.09
ATOM	991	CA	GLY A	143	13.210	14.075	14.757		11.80
ATOM	992	С	GLY A	143	12.203	12.972	14.555		16.69
MOTA	993	0	GLY A	143	11.760	12.787	13.481		19.57
ATOM	994	N	ILE A	144	11.668	12.386	15.590		19.71
MOTA	995	CA	ILE A	144	10.494	11.589	15.667		20.13
MOTA	996	С	ILE A	144	9.313	12.315	16.296	1.00	
MOTA	997	0	ILE A	144	9.298	13.026	17.268	1.00	
MOTA	998	CB	ILE A	144	10.973	10.583	16.692	1.00	
MOTA	999	CG1	ILE A	144	12.363	9.956	16.348	1.00	5.60
MOTA	1000	CG2	ILE A	144	9.882	9.636	16.775		14.01
MOTA	1001	CD1	ILE A	144	12.437	9.156	17.562	1.00	2.75
ATOM	1002	N	PRO A	145	8.249	12.380	15.499		32.77
MOTA	1003	CA	PRO A	145	6.959	12.993	15.779	1.00	
ATOM	1004	С	PRO A	145	6.484	12.588	17.180	1.00	
ATOM	1005	0	PRO A	145	6.475	11.446	17.537	1.00	
MOTA	1006	CB	PRO A	145	5.957	12.384	14.784	1.00	
ATOM	1007	CG	PRO A	145	6.887	12.059	13.668	1.00	

ATOM	1008	CD	PRO	Α	145		8.174	11.563	14.234	1.00	31.33
ATOM	1009	N	ASN	A	146		5.796	13.462	17.878	1.00	27.07
ATOM	1010	CA	ASN	Α	146		5.454	13.274	19.230	1.00	28.59
MOTA	1011	С			146		6.526	12.605	20.045	1.00	29.25
ATOM	1012	0	ASN	Α	146		6.087	11.995	20.996	1.00	35.51
ATOM	1013	CB	ASN	Α	146		4.285	12.364	19.230	1.00	41.13
ATOM	1014	CG	ASN	Α	146		3.300	12.568	18.120	1.00	48.43
ATOM	1015	OD1	ASN	Α	146		3.134	13.721	17.788	1.00	49.24
ATOM	1016	ND2	ASN	Α	146		2.763	11.437	17.695	1.00	47.79
MOTA	1017	N	TYR	Α	147		7.791	12.799	19.885	1.00	23.88
MOTA	1018	CA	TYR	Α	147	•	8.689	12.339	20.969	1.00	21.90
ATOM	1019	С	TYR	Α	147		9.583	13.495	21.285	1.00	22.57
ATOM	1020	0	TYR	Α	147		9.777	14.399	20.494	1.00	26.53
ATOM	1021	CB	TYR	Α	147		9.309	11.098	20.498	1.00	21.16
MOTA	1022	CG	TYR	Α	147	1	0.285	10.471	21.349	1.00	20.45
ATOM	1023	CD1	TYR	A	147		9.882	9.720	22.384	1.00	24.28
ATOM	1024	CD2	TYR	Α	147	1	1.608	10.564	21.189	1.00	17.96
ATOM	1025	CE1		Α	147	1	0.681	9.029	23.273	1.00	24.55
ATOM	1026	CE2	TYR	Α	147	1	2.509	9.948	21.983	1.00	20.73
ATOM	1027	CZ	TYR	A	147	1	2.022	9.184	23.030	1.00	24.61
ATOM	1028	OH	TYR	Α	147	1	2.891	8.536	23.887	1.00	24.80
MOTA	1029	N	PRO	Α	148		9.893	13.858	22.507	1.00	22.86
ATOM	1030	CA	PRO			1	0.817	14.916	22.769	1.00	21.77
ATOM	1031	С	PRO	A	148	1	2.127	14.882	21.957	1.00	22.49
MOTA	1032	0	PRO			1	3.007	14.004	22.117	1.00	22.31
ATOM	1033	CB	PRO			1	1.185	14.694	24.251	1.00	23.23
ATOM	1034	CG	PRO	Α	148	1	0.324	13.576	24.719	1.00	23.39
ATOM	1035	CD	PRO				9.677	12.889	23.590	1.00	25.33
ATOM	1036	N	ARG			1	2.432	15.980	21.250	1.00	25.45
ATOM	1037	CA	ARG			1	3.735	16.138	20.567	1.00	22.54
ATOM	1038	С	ARG			1	4.910	16.018	21.499	1.00	21.28
ATOM	1039	0	ARG			1	5.860	15.477	21.015	1.00	16.61
MOTA	1040	CB	ARG			1	3.829	17.346	19.727	1.00	31.02
MOTA	1041	CG	ARG			1	2.837	17.750	18.719	1.00	58.26
ATOM	1042	CD	ARG			1	3.452	18.605	17.658	1.00	80.58
ATOM	1043	NE	ARG			1	3.769	17.798	16.491	1.00	92.05
MOTA	1044	CZ	ARG			1	3.315	18.154	15.320	1.00	91.85
ATOM	1045		ARG			1	2.586	19.213	15.165	1.00	86.98
MOTA	1046	NH2	ARG	Α	149	1	3.544	17.488	14.242	1.00	91.61
MOTA	1047	N	GLU			1	4.813	16.282	22.825	1.00	28.09
MOTA	1048	CA	GLU			1	5.950	16.171	23.735	1.00	25.55
ATOM	1049	С	GLU			1	6.272	14.736	24.020		21.12
ATOM	1050	0	GLU				7.372	14.443	24.371	1.00	24.39
ATOM	1051	CB	GLU				5.753	17.040	24.917	1.00	38.73
ATOM	1052	CG	GLU			1	4.328	17.370	25.359	1.00	67.27
ATOM	1053	CD	GLU				4.252	17.185	26.899		85.05
ATOM	1054		GLU				5.005	17.890	27.657	1.00	90.70
ATOM	1055	OE2	GLU	A	150	1	3.454	16.321	27.373	1.00	91.68

ATOM	1056	N	ARG	Α	151	15.396	13.807	23.727	1.00	19.70
ATOM	1057	CA	ARG	Α	151	15.752	12.424	23.844		19.52
ATOM	1058	С	ARG	Α	151	16.163	11.779	22.531		19.28
ATOM	1059	0	ARG	Α	151	16.373	10.586	22.480		14.55
ATOM	1060	CB			151	14.548	11.796	24.412	1.00	
ATOM	1061	CG			151	13.853	12.432	25.516	1.00	
ATOM	1062	CD			151	13.200	11.451	26.393	1.00	
ATOM	1063	NE			151	12.609	11.893	27.633	1.00	
ATOM	1064	CZ	ARG			11.796	11.028	28.275		52.87
ATOM	1065	NH1				11.428	9.823	27.930		51.02
ATOM	1066		ARG			11.203	11.278	29.416		59.98
ATOM	1067	N	THR			16.360	12.526			
ATOM	1068	CA	THR			16.629		21.505	1.00	
ATOM	1069	C	THR				11.925	20.253	1.00	
ATOM	1070	0	THR			17.995	12.249	19.745		17.30
ATOM	1070	CB				18.282	13.373	19.965		21.34
ATOM	1071	OG1	THR			15.680	12.408	19.158		13.91
	1072					14.423	12.256	19.858	1.00	
ATOM ATOM	1073	CG2				15.737	11.934	17.759	1.00	6.77
	1074	N	LYS			18.704	11.336	19.121		15.49
ATOM		CA	LYS			19.930	11.725	18.450	1.00	17.73
ATOM	1076	С	LYS			19.893	11.035	17.073		18.41
ATOM	1077	0	LYS			19.866	9.800	17.121		16.04
ATOM	1078	CB	LYS			21.112	11.260	19.338		14.55
ATOM	1079	CG	LYS			22.523	11.508	18.933		11.95
ATOM	1080	CD	LYS			22.883	12.882	19.403	1.00	40.35
ATOM	1081	CE	LYS			24.358	13.093	19.079	1.00	62.12
ATOM	1082	NZ	LYS			24.930	14.235	19.863	1.00	73.03
ATOM	1083	N	VAL			19.910	11.962	16.136	1.00	15.86
MOTA	1084	CA	VAL			20.031	11.508	14.730	1.00	15.79
MOTA	1085	С	VAL			21.406	11.481	14.040	1.00	13.11
ATOM	1086	0	VAL			21.958	12.460	13.675	1.00	13.51
ATOM	1087	CB	VAL			19.095	12.257	13.674	1.00	5.90
MOTA	1088	CG1	VAL	A	154	19.276	11.765	12.247	1.00	8.45
MOTA	1089	CG2	VAL	A	154	17.672	12.091	14.117	1.00	7.14
ATOM	1090	N	PHE	A	155	22.039	10.448	13.605	1.00	13.75
MOTA	1091	CA	PHE	Α	155	23.263	10.473	12.843	1.00	10.67
MOTA	1092	С	PHE	Α	155	22.906	10.406	11.402		11.64
MOTA	1093	0	PHE	A	155	22.505	9.367	10.893		15.09
ATOM	1094	СВ	PHE	A	155	23.955	9.120	13.304	1.00	5.38
ATOM	1095	CG	PHE	Α	155	24.396	9.266	14.739		16.52
MOTA	1096		PHE			23.678	8.642	15.696		23.70
ATOM	1097		PHE			25.503	9.950	15.107		11.27
ATOM	1098	CE1	PHE			24.037	8.702	17.011		23.25
ATOM	1099		PHE			25.888	9.994	16.372		7.37
ATOM	1100	CZ	PHE			25.139	9.384	17.357		16.13
MOTA	1101	N	CYS			23.205	11.255	10.511		12.38
ATOM	1102	CA	CYS			22.847	11.233	9.114		11.64
ATOM	1103	C	CYS			24.057	12.027	8.461		
		_	UIU .		100	23.00/	12.02/	0.401	1.00	10.08

ATOM	1104	0	CYS			.385	13.174	8.378	1.00	13.73
MOTA	1105	CB	CYS			.575	12.391	8.917	1.00	6.30
MOTA	1106	SG	CYS :	A 15	6 20	.137	11.470	8.287	1.00	10.60
ATOM	1107	N	ASN .	A 15	7 24	.814	11.147	7.918	1.00	16.95
ATOM	1108	CA	ASN .	A 15	7 26	.229	11.665	7.576	1.00	19.16
ATOM	1109	С	ASN :	A 15	7 26	.197	12.367	6.310	1.00	17.70
ATOM	1110	0	ASN I	A 15	7 25	.368	12.330	5.469	1.00	20.91
ATOM	1111	CB	ASN I	A 15			10.714	8.300		30.34
ATOM	1112	CG	ASN 3	A 15		.733	9.498	7.932		34.95
ATOM	1113	OD1	ASN I	A 15		.011	8.573	8.606		44.28
ATOM	1114		ASN I			.965	9.541	6.660		54.18
ATOM	1115	N	VAL I				13.501	6.313		25.65
ATOM	1116	CA	VAL I				14.483	5.192		28.21
ATOM	1117	С	VAL Z				13.893	3.758		24.85
ATOM	1118	ō	VAL Z				14.266	3.111		30.96
ATOM	1119	CB	VAL 2				15.512	5.217		27.87
ATOM	1120	CG1	VAL 2				14.595	4.238		40.51
ATOM	1121	CG2	VAL A				16.704	4.399		34.39
ATOM	1122	N	GLY A				12.956			5.94
ATOM	1123	CA	GLY A				12.774	3.016	1.00	
ATOM	1124	C	GLY A					1.732	1.00	6.20
ATOM	1125	0	GLY A				11.797	1.487	1.00	4.00
ATOM	1126	N	ASP A				10.704	0.848	1.00	4.06
ATOM	1127	CA	ASP A				11.441	2.643	1.00	8.53
ATOM							10.302	2.828		11.97
	1128	C	ASP A				10.698	2.177		14.44
ATOM	1129	0	ASP A				11.398	2.692		10.21
ATOM	1130	CB	ASP A			.037	9.829	4.277		12.43
ATOM	1131	CG	ASP A			.126	8.629	4.261		20.99
ATOM	1132	OD1	ASP A		_	.525	8.408	3.179		33.03
MOTA	1133	OD2	ASP A			.956	7.840	5.216		10.13
ATOM	1134	N	ALA A				10.402	0.961		12.33
ATOM	1135	CA	ALA A				10.743	0.269		11.01
MOTA	1136	С	ALA A				10.317	0.848	1.00	15.22
MOTA	1137	0	ALA A				11.034	0.594	1.00	9.50
MOTA	1138	CB	ALA A				10.334	-1.172	1.00	13.68
MOTA	1139	N	VAL A			.915	9.468	1.840	1.00	14.54
ATOM	1140	CA.	VAL A			. 653	9.014	2.287	1.00	9.86
ATOM	1141	С	VAL A			.235	10.063	3.258	1.00	13.50
MOTA	1142	0	VAL A	16	2 17.	.094	10.458	3.377	1.00	20.47
MOTA	1143	CB	VAL A	16	2 18.	.596	7.778	3.117	1.00	7.34
MOTA	1144	CG1	VAL A	16	2 18.	.931	6.592	2.259	1.00	6.50
MOTA	1145	CG2	VAL A	16	2 19.	.514	7.858	4.210		18.46
MOTA	1146	N	CYS A	16			10.733	3.719		13.44
MOTA	1147	CA	CYS A	16			11.811	4.720		11.26
MOTA	1148	С	CYS A	16			12.963	4.042		15.57
MOTA	1149	0	CYS A	16			13.857	4.880		14.09
MOTA	1150	CB	CYS A	16			12.145	5.570		18.70
MOTA	1151	SG	CYS A	16			10.705	6.581		13.38

MOTA	1152	N	THR	Α	164	18.100	13.014	2.696	1.00	21.82
MOTA	1153	CA			164	17.603	14.283	2.171	1.00	23.08
ATOM	1154	С	THR	A	164	16.597	14.022	1.098	1.00	23.39
MOTA	1155	0	THR	Α	164	16.517	14.727	0.137	1.00	33.37
MOTA	1156	CB	THR	Α	164	18.463	15.341	1.454	1.00	23.25
MOTA	1157	OG1	THR	Α	164	19.486	14.707	0.674	1.00	23.21
ATOM	1158	CG2	THR	Α	164	18.958	16.261	2.491	1.00	37.71
MOTA	1159	N	GLY	Α	165	15.802	13.085	1.309	1.00	24.23
ATOM	1160	CA	GLY	A	165	14.606	12.783	0.579	1.00	26.69
MOTA	1161	С	GLY	Α	165	14.699	11.814	-0.515	1.00	28.56
ATOM	1162	0	GLY	Α	165	13.680	11.775	-1.124	1.00	39.76
ATOM	1163	N	THR	Α	166	15.661	11.044	-0.736	1.00	25.80
ATOM	1164	CA	THR	Α	166	16.006	10.220	-1.774	1.00	25.53
ATOM	1165	С	THR	Α	166	16.195	8.866	-1.175	1.00	25.35
ATOM	1166	0	THR	Α	166	16.913	8.760	-0.206	1.00	30.91
ATOM	1167	CB	THR	Α	166	17.406	10.657	-2.230	1.00	31.57
ATOM	1168	OG1	THR	Α	166	17.105	11.788	-2.982	1.00	24.13
ATOM	1169	CG2	THR	Α	166	18.061	9.559	-2.983	1.00	34.67
ATOM	1170	N	LEU	Α	167	15.734	7.833	-1.817	1.00	19.63
ATOM	1171	CA	LEU	Α	167	16.219	6.552	-1.465	1.00	16.11
ATOM	1172	C	LEU	A	167	17.395	6.044	-2.300	1.00	19.87
ATOM	1173	0	LEU	A	167	17.265	4.869	-2.612	1.00	21.38
ATOM	1174	CB	LEU	Α	167	15.086	5.624	-1.555	1.00	23.45
ATOM	1175	CG	LEU	Α	167	14.123	5.773	-0.401	1.00	33.91
ATOM	1176	CD1	LEU	Α	167	12.969	4.908	-0.793	1.00	42.10
ATOM	1177	CD2	LEU	A	167	14.776	5.385	0.903	1.00	25.86
ATOM	1178	N	ILE	Α	168	18.534	6.726	-2.507	1.00	21.67
ATOM	1179	CA	ILE	Α	168	19.608	6.051	-3.170	1.00	23.38
ATOM	1180	С	ILE	Α	168	20.675	5.585	-2.189	1.00	20.47
ATOM	1181	0	ILE	Α	168	21.139	6.541	-1.581	1.00	18.08
ATOM	1182	CB	ILE	Α	168	20.254	6.835	-4.297	1.00	
ATOM	1183	CG1	ILE	A	168	21.232	7.874	-3.800	1.00	
ATOM	1184	CG2	ILE	Α	168	19.445	7.627	-5.276	1.00	
ATOM	1185	CD1	ILE	Α	168	20.908	8.938	-4.804		26.95
MOTA	1186	N	ILE	Α	169	21.396	4.478	-2.394	1.00	18.32
ATOM	1187	CA	ILE	A	169	22.554	4.448	-1.536		13.25
ATOM	1188	С	ILE	Α	169	23.924	4.662	-1.967	1.00	11.95
ATOM	1189	0	ILE	Α	169	24.615	3.942	-2.539	1.00	20.35
ATOM	1190	CB	ILE	Α	169	22.503		-0.499		21.07
ATOM	1191	CG1	ILE	A	169	23.398		-0.655		11.06
ATOM	1192	CG2	ILE	Α	169	21.122	2.801	-0.533		7.02
ATOM	1193	CD1	ILE	A	169	22.581	1.266	-1.587		32.83
ATOM	1194	N	THR	Α	170	24.570	5.586	-1.296		17.16
ATOM	1195	CA	THR	Α	170	25.883		-1.397		13.01
ATOM	1196	С	THR	A	170	26.722		-0.240		10.14
ATOM	1197	0	THR			26.334		0.758	1.00	
ATOM	1198	CB	THR	A	170	25.623				15.02
ATOM	1199	OG1	THR	Α	170	26.466	7.947	-0.255		23.39

ATOM	1200	CG2			170	24.389	7.914	-0.452	1.00	41.10
ATOM	1201	N			171	28.000	5.738	-0.469	1.00	10.12
ATOM	1202	CA			171	29.012	5.066	0.339	1.00	11.88
ATOM	1203	С			171	28.897	5.492	1.765	1.00	9.74
ATOM	1204	0	PRO	Α	171	28.904	4.682	2.646	1.00	9.54
ATOM	1205	CB	PRO	Α	171	30.414	5.207	-0.286	1.00	7.15
ATOM	1206	CG	PRO	Α	171	30.017	5.603	-1.654	1.00	7.18
ATOM	1207	CD	PRO	Α	171	28.667	6.233	-1.601	1.00	6.90
ATOM	1208	N	ALA	Α	172	28.725	6.718	1.980	1.00	6.71
ATOM	1209	CA	ALA	Α	172	28.247	7.315	3.169	1.00	8.62
ATOM	1210	С	ALA	Α	172	27.075	6.631	3.892	1.00	
ATOM	1211	0	ALA	A	172	27.037	6.755	5.165		16.49
ATOM	1212	CB	ALA	Α	172	27.904	8.812	3.040	1.00	2.86
ATOM	1213	N	HIS	Α	173	26.287	5.815	3.278	1.00	6.36
ATOM	1214	CA	HIS	Α	173	25.133	5.468	4.081	1.00	5.29
ATOM	1215	С	HIS	Α	173	25.685	4.314	4.888		10.58
ATOM	1216	0	HIS	Α	173	25.082	3.598	5.668	1.00	9.36
ATOM	1217	CB	HIS	Α	173	24.081	4.883	3.216	1.00	8.41
ATOM	1218	CG	HIS	Α	173	22.815	4.403	3.791	1.00	7.30
MOTA	1219	ND1	HIS	Α	173	22.066	5.327	4.565	1.00	8.48
ATOM	1220		HIS			22.148	3.264	3.670	1.00	7.83
ATOM	1221		HIS			20.932	4.657	4.861	1.00	17.36
ATOM	1222	NE2	HIS	Α	173	20.945	3.423	4.379	1.00	5.29
ATOM	1223	N	LEU	Α	174	26.823	3.947	4.326	1.00	8.03
ATOM	1224	CA	LEU	Α	174	27.344	2.623	4.682	1.00	8.06
ATOM	1225	С	LEU	Α	174	28.171	2.787	5.930	1.00	13.06
ATOM	1226	0	LEU	Α	174	28.609	1.648	6.151	1.00	19.88
ATOM	1227	CB	LEU	Α	174	28.078	2.118	3.488	1.00	2.76
MOTA	1228	CG	LEU	A	174	27.560	0.902	2.847	1.00	13.35
ATOM	1229	CD1	LEU	A	174	26.024	1.017	2.796	1.00	18.01
MOTA	1230	CD2	LEU	Α	174	27.913	0.740	1.421	1.00	21.70
ATOM	1231	N	SER			28.290	3.989	6.447	1.00	12.43
ATOM	1232	CA	SER	Α	175	29.230	4.052	7.553	1.00	18.01
ATOM	1233	С	SER	A	175	28.872	4.811	8.847	1.00	19.89
ATOM	1234	0	SER	A	175	28.968	6.047	9.120	1.00	14.61
ATOM	1235	CB	SER			30.516	4.606	6.847	1.00	20.11
ATOM	1236	OG	SER			30.834	5.907	7.293	1.00	27.73
ATOM	1237	N	TYR			28.479	3.978	9.815	1.00	17.89
ATOM	1238	CA	TYR			28.092	4.530	11.133	1.00	12.54
ATOM	1239	С	TYR			28.530	3.671	12.272	1.00	11.16
ATOM	1240	0	TYR			27.949	3.770	13.257	1.00	7.63
ATOM	1241	CB	TYR			26.511	4.283	11.053	1.00	9.13
ATOM	1242	CG	TYR			25.831	5.525	10.029	1.00	5.03
ATOM	1243	CD1	TYR			25.874	6.923	10.425	1.00	2.75
ATOM	1244	CD2	TYR			25.152	5.022	8.980	1.00	2.18
ATOM	1245	CE1	TYR			25.287	7.754	9.633	1.00	4.25
ATOM	1246	CE2	TYR			24.649	5.981	8.085	1.00	6.77
MOTA	1247	CZ	TYR	A	176	24.658	7.329	8.399	1.00	6.22

ATOM	1248	OH	TYR	. A	176	24.	074	8.375	7.635	1.00	5.76
ATOM	1249	N	THR	A	177		430	2.685	12.167		
ATOM	1250	CA	THR	A	177	29.	797	1.854	13.284	1.00	
ATOM	1251	С	THR	A	177	30.	516	2.659	14.320	1.00	12.46
ATOM	1252	0	THR	A	177	30.	311	2.436	15.475	1.00	13.12
ATOM	1253	CB	THR	A	177	30.	658	0.683	12.798		
ATOM	1254	OG1	THR	A	177	31.	361	1.247	11.870	1.00	
MOTA	1255	CG2	THR	A	177	29.	675	-0.149	12.083	1.00	6.42
MOTA	1256	N	ILE	A	178	31.	409	3.474	13.920	1.00	10.48
ATOM	1257	CA	ILE	Α	178	32.	203	4.246	14.783	1.00	15.25
ATOM	1258	С	ILE	A	178	31.	180	5.045	15.632		16.95
ATOM	1259	0	ILE	A	178	31.	092	4.774	16.851		22.68
ATOM	1260	CB	ILE	A	178	33.	338	5.121	14.357		25.11
ATOM	1261	CG1	ILE	A	178	34.	701	4.496	14.056		25.05
ATOM	1262	CG2	ILE	A.	178	33.	599	6.205	15.392		27.60
ATOM	1263	CD1	ILE	A	178		553	3.006	14.071		55.86
ATOM	1264	N	GLU	A	179		218	5.799	15.178		16.34
ATOM	1265	CA	GLU	A	179		290	6.610	15.985		16.94
ATOM	1266	С	GLU	Α	179		324	5.713	16.692		14.79
ATOM	1267	0	GLU	A	179		683	6.012	17.716		19.20
ATOM	1268	CB	GLU	A	179		555	7.637	15.169		21.16
ATOM	1269	CG	GLU	A	179	28.	790	7.283	13.691		50.37
ATOM	1270	CD	GLU	Α	179	29.	933	7.701	12.851		61.82
ATOM	1271	OE1	GLU	A	179	30.	163	8.890	12.697		77.56
ATOM	1272	OE2	GLU	A	179	30.	627	6.854	12.309		75.83
ATOM	1273	N	ALA	A	180	28.	240	4.418	16.412	1.00	8.00
ATOM	1274	CA	ALA	Α	180	27.	353	3.520	17.042	1.00	14.34
ATOM	1275	С	ALA	Α	180	28.	048	2.991	18.280	1.00	19.53
ATOM	1276	0	ALA	A	180	27.	397	3.142	19.265		21.17
ATOM	1277	CB	ALA	Α	180	26.	843	2.437	16.128		11.97
ATOM	1278	N	ARG	A	181	29.	317	2.547	18.287		21.89
ATOM	1279	CA	ARG	A	181	29.	992	1.982	19.398		16.48
ATOM	1280	С	ARG	Α	181	30.	296	3.106	20.367		19.44
ATOM	1281	0	ARG	Α	181	30.	243	3.104	21.639	1.00	28.53
ATOM	1282	CB	ARG	Α	181	31.	310	1.408	19.143		12.43
ATOM	1283	CG	ARG	Α	181	31.	954	0.432	20.052		45.44
ATOM	1284	CD	ARG	Α	181	32.	596	-0.688	19.242	1.00	66.21
ATOM	1285	NE	ARG	Α	181	33.	333	-0.030	18.164		85.83
ATOM	1286	CZ	ARG	Α	181	33.	306	-0.321	16.895		91.35
ATOM	1287		ARG			32.		-1.320	16.530	1.00	96.98
ATOM	1288	NH2	ARG	Α	181	34.	023	0.400	16.095		92.83
ATOM	1289	N	GLY	Α	182	30.		4.262	19.847	1.00	13.94
MOTA	1290	CA	GLY	A	182	30.		5.404	20.728	1.00	7.40
ATOM	1291	С	GLY	Α	182	29.		6.574	20.960	1.00	7.95
ATOM	1292	0	GLY			29.	171 -	6.512	22.083	1.00	12.73
ATOM	1293	N	GLU	A	183	29.		7.622	20.138	1.00	6.42
ATOM	1294	CA	GLU	A	183	28.		8.775	20.405	1.00	10.04
ATOM	1295	С	GLU	Α	183	27.	421	8.369	20.645	1.00	14.41
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ATOM	1296	0	GLU	Α	183	26.846	8.530	21.749	1.00	15.43
ATOM	1297	CB	GLU	Α	183	29.053	9.791	19.402		21.24
ATOM	1298	CG	GLU	A	183	28.079	10.638	18.725	1.00	
MOTA	1299	CD	GLU	Α	183	28.248	12.103	19.141	1.00	
ATOM	1300	OE1	GLU	Α	183	28.850	12.243	20.232	1.00	
ATOM	1301	OE2			183	27.791	13.027	18.430	1.00	
ATOM	1302	N			184	26.766	7.605	19.808	1.00	
MOTA	1303	CA			184	25.444	7.083	20.117	1.00	
ATOM	1304	С			184	25.549	6.382	21.464		13.62
ATOM	1305	0			184	24.575	6.533	22.215	1.00	
ATOM	1306	CB			184	25.019	6.015	19.089	1.00	9.58
ATOM	1307	N			185	26.428	5.396	21.774	1.00	9.42
MOTA	1308	CA			185	26.219	4.677	23.031	1.00	7.48
ATOM	1309	С			185	26.330	5.715	24.100	1.00	
ATOM	1310	0			185	25.761	5.503	25.179	1.00	9.50
ATOM	1311	CB			185	27.138	3.475	23.260	1.00	4.60
ATOM	1312	N			186	27.271	6.673	24.090	1.00	
ATOM	1313	CA	ARG			27.352	7.507	25.300	1.00	
ATOM	1314	С	ARG	Α	186	26.085	8.286	25.561	1.00	
ATOM	1315	0	ARG	Α	186	25.421	8.267	26.573	1.00	8.74
ATOM	1316	CB	ARG	Α	186	28.484	8.460	25.043	1.00	
MOTA	1317	CG	ARG	A	186	29.869	7.851	25.240		37.15
MOTA	1318	CD	ARG	Α	186	30.983	8.826	24.813	1.00	
ATOM	1319	NE	ARG	Α	186	31.902	7.942	24.064		51.82
MOTA	1320	CZ	ARG	Α	186	32.324	8.346	22.870	1.00	50.20
MOTA	1321	NH1	ARG	Α	186	31.924	9.538	22.424		47.65
ATOM	1322	NH2	ARG	Α	186	33.115	7.476	22.318	1.00	39.90
ATOM	1323	N	PHE	Α	187	25.565	8.774	24.434	1.00	8.37
ATOM	1324	CA	PHE	A	187	24.195	9.370	24.426	1.00	13.48
ATOM	1325	С	PHE	A	187	23.187	8.476	25.182	1.00	15.92
MOTA	1326	0	PHE		187	22.379	8.916	25.995	1.00	14.81
ATOM	1327	CB	PHE	Α	187	23.667	9.791	23.087		11.81
ATOM	1328	CG	PHE	A	187	22.282	10.323	23.032	1.00	14.64
ATOM	1329	CD1	PHE	Α	187	21.984	11.586	23.391	1.00	8.47
ATOM	1330	CD2			187	21.186	9.599	22.564	1.00	18.34
ATOM	1331	CE1			187	20.698	12.134	23.353	1.00	12.89
ATOM	1332	CE2	PHE	A	187	19.895	10.026	22.485	1.00	17.42
ATOM	1333	CZ	PHE	Α	187	19.661	11.322	22.924	1.00	
ATOM	1334	N	LEU			23.033	7.232	24.803	1.00	
ATOM	1335	CA	LEU	A	188	21.908	6.427	25.324	1.00	
ATOM	1336	С	LEU			22.207	6.221	26.775	1.00	
ATOM	1337	0	LEU			21.280	6.512	27.461	1.00	18.17
ATOM	1338	CB	LEU			21.703	5.088	24.552	1.00	18.72
ATOM	1339	CG	LEU			21.116	5.375	23.136	1.00	
ATOM	1340		LEU			20.950	4.066	22.601	1.00	7.86
ATOM	1341		LEU			19.849	6.206	23.168	1.00	4.70
ATOM	1342	N	ARG			23.333	5.805	27.230	1.00	17.48
ATOM	1343	CA	ARG	A	189	23.798	5.812	28.547	1.00	18.41

ATOM	1344	С	ARG	А	189		23.353	7.039	29.321	1.00	16.87
ATOM	1345	0	ARG	Α	189		22.852	7.164	30.389	1.00	13.64
ATOM	1346	CB	ARG	Α	189		25.325	6.017	28.529	1.00	21.93
ATOM	1347	CG	ARG	Α	189		25.882	5.624	29.894	1.00	19.95
ATOM	1348	CD ·	ARG	Α	189		27.239	6.140	30.235		21.42
ATOM	1349	NE	ARG	Α	189		27.257	7.545	29.926		25.62
ATOM	1350	CZ	ARG	Α	189		28.491	7.983	29.699	1.00	
ATOM	1351	NH1	ARG	Α	189		29.315	6.960	29.840		26.71
ATOM	1352	NH2	ARG	Α	189		28.780	9.210	29.383	1.00	
ATOM	1353	N	ASP	Α	190		23.837	8.150	28.796	1.00	
ATOM	1354	CA	ASP	Α	190	•	23.489	9.338	29.615		17.78
ATOM	1355	С	ASP	Α	190		22.008	9.364	29.711	1.00	
ATOM	1356	0	ASP	Α	190		21.661	9.891	30.692		23.13
ATOM	1357	CB	ASP	Α	190		23.995	10.663	29.070		23.17
ATOM	1358	CG	ASP	Α	190		25.553	10.664	29.079	1.00	
ATOM	1359	OD1	ASP	Α	190		26.250	9.836	29.761		22.68
ATOM	1360	OD2					25.961	11.595	28.321	1.00	
ATOM	1361	N	ARG	A	191		21.156	9.128	28.781		21.61
ATOM	1362	CA	ARG	Α	191		19.707	9.265	28.849	1.00	
ATOM	1363	С	ARG	Α	191		19.176	8.237	29.825		21.23
ATOM	1364	0	ARG	Α	191		18.327	8.515	30.651	1.00	
ATOM	1365	CB	ARG .	Α	191		19.014	9.214	27.450	1.00	19.76
ATOM	1366	CG	ARG .	Α	191		19.605	10.282	26.521		27.49
ATOM	1367	CD	ARG .				18.848	11.594	26.689	1.00	
ATOM	1368	NE	ARG .	Α	191		17.559	11.023	27.144		60.89
ATOM	1369	CZ	ARG .				16.841	11.651	28.087		73.30
ATOM	1370	NH1	ARG .				17.404	12.780	28.496		76.65
ATOM	1371	NH2	ARG .				15.675	11.224	28.574		62.02
ATOM	1372	N	ILE .				19.734	7.037	29.885		21.02
ATOM	1373	CA	ILE :				19.500	6.080	30.913		21.92
ATOM	1374	С	ILE :				19.705	6.598	32.337		25.67
ATOM	1375	0	ILE :				19.145	6.053	33.263		27.95
ATOM	1376	CB	ILE :				20.289	4.775	30.750		24.23
ATOM	1377	CG1	ILE				19.770	4.215	29.475		26.91
ATOM	1378	CG2	ILE :				19.923	3.983	31.951		15.15
ATOM	1379	CD1	ILE :				20.418	2.954	29.019		21.07
ATOM	1380	N	ARG :				20.535	7.574	32.629		28.72
ATOM	1381	CA	ARG 2	A	193		20.800	8.068	33.963		33.95
ATOM	1382	С	ARG I				20.116	9.377	34.406		42.87
ATOM	1383	0	ARG :				20.479	9.267	35.618		48.19
ATOM	1384	CB	ARG :				22.298	8.179	34.167		34.19
ATOM	1385	CG	ARG I				23.096	6.896	34.100		39.38
ATOM	1386	CD	ARG 2				24.590	7.213	34.133		65.92
ATOM	1387	NE	ARG Z			•	25.339	5.973	34.003		81.05
ATOM	1388	CZ	ARG A				26.631	5.765	33.770	1.00	
ATOM	1389		ARG A				27.441	6.816	33.647	1.00	
ATOM	1390		ARG 2				27.120	4.536	33.652	1.00	
ATOM	1391	OT	ARG A				19.292	10.277	34.082	1.00	
TER		_		-					31.002	1.00	50.00

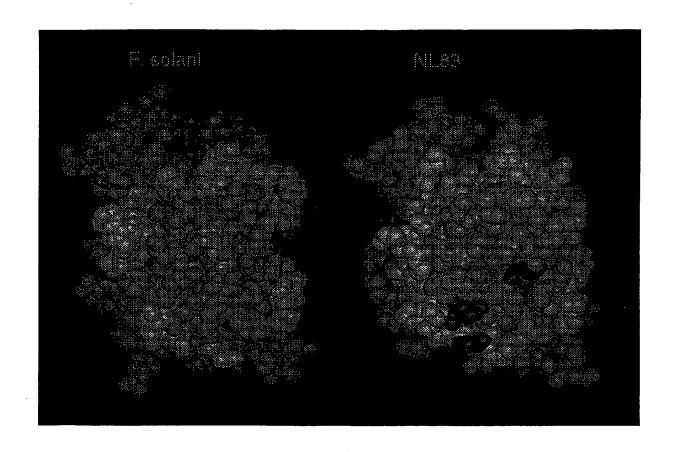


Fig. 2
3D structure of cutinases from *F. solani pisi* (left) and *H. insolens* (right)

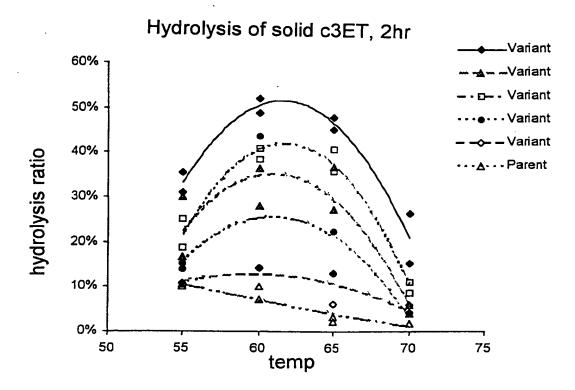


Fig. 3 Hydrolysis of solid c3ET, 2 hr

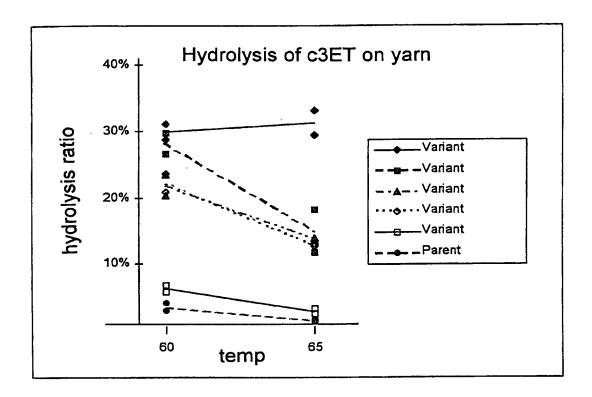


Fig. 4
Hydrolysis of c3ET on yarn, 17 hr

WO 00/34450

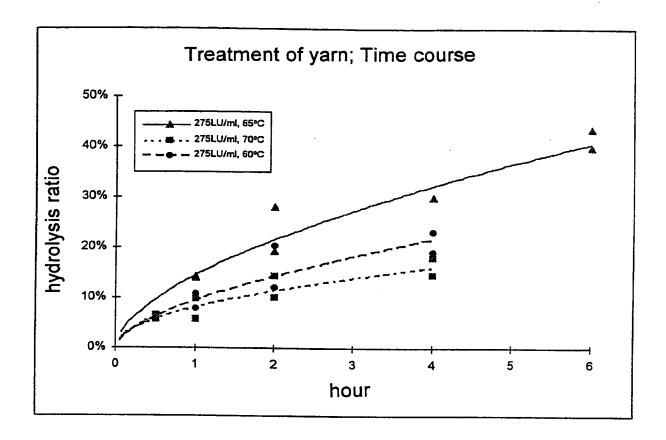


Fig. 5
Treatment of yarn; time course

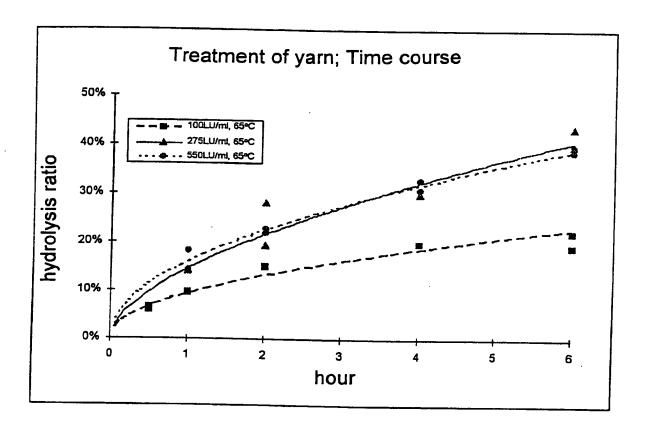


Fig. 6
Treatment of yarn; time course

International application No.

PCT/DK 99/00678

A. CLASSIFICATION OF SUBJECT MATTER

IPC7: C12N 9/18 // C11D 3/386, C08G 63/91
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC7: C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE, DK, FI, NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

C.	DOCUMENTS	CONSIDERED	TO	BE	RELEVANT	
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Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 9009446 A1 (PLANT GENETICS SYSTEMS, N.V.), 23 August 1990 (23.08.90), see page 1, lines 11-20, claims	1-32,34
		į
x	WO 9414963 A1 (UNILEVER N.V.), 7 July 1994 (07.07.94), see claim 14	1-32,34
A	WO 9414964 AI (UNILEVER N.V.), 7 July 1994 (07.07.94)	1-32,34
		
A	WO 9704078 A1 (NOVO NORDISK A/S), 6 February 1997 (06.02.97), see claim 51	1-32,34

<u> </u>						
X	Further documents are listed in the continuation of Box	с С.	X See patent family annex.			
* "A"	Special categories of cited documents: document defining the general state of the art which is not considered to be of particular relevance	Т.	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention			
"E"	erlier document but published on or after the international filing date document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other	"X"	document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone			
"O"	special reason (as specified) O" document referring to an oral disclosure, use, exhibition or other means		"document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art			
Date	the priority date claimed e of the actual completion of the international search	"& "	document member of the same patent family			
	May 2000	Date	of mailing of the international search report 1 1 -05- 2000			
Swe	ne and mailing address of the ISA/ edish Patent Office		rized officer			
	: 5055, S-102 42 STOCKHOLM simile No. + 46 8 666 02 86		nne Siösteen/EÖ none No + 46 8 782 25 00			

Form PCT/ISA/210 (second sheet) (July 1992)

International application No.
PCT/DK 99/00678

C (Continu	ation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*		vant nassagge	Relevant to claim No
Jalegory	Chauton of document, with indication, where appropriate, of the rele	vant passages	Relevant to claim No
A	PROTEINS: Structure, Function, and Genetics, Volume 26, 1996, Sonia Longhi et al, "Dyn Fusarium solani Cutinase Investigated Thr Structural Comparison Among Different Cry Forms of Its Variants" page 442 - page 45	amics of ough stal 8	1-32,34
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	A/210 (continuation of second sheet) (July 1992)		

International application No. PCT/DK 99/00678

Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This inte	ernational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1.	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2.	Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).:
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
	ernational Searching Authority found multiple inventions in this international application, as follows: next sheet
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-32 and 34
Remark o	on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

International application No. PCT/DK 99/00678

The invention claimed relates to two different inventions :

- I. Claims 1-32 and 34 relate to cutinase variants and the use of these variants.
- II. Claim 33 relates to a method for detecting cutinase activity in a sample.

Unity of invention exists only when there is a technical relationship among the claimed inventions involving one or more of the same or corresponding "special technical feature" i.e. features that define a contribution which each of the inventions make over prior art. (See Annex B to administrative instructions and Rule 13.1).